

**Summary Report**

**A Study of Macroinvertebrate Community Responses  
to Winter Flows on the Fryingpan River**

**Prepared for**

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**August 11, 2004**



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## **INTRODUCTION**

The biological and physical processes that govern the structure and function of benthic macroinvertebrate communities in the Fryingpan River are not entirely understood. However, there is evidence to suggest that the flow regime may be an important physical influence on benthic communities (Rees et al. 2003). For this reason, macroinvertebrate sampling and thermal modeling continued during the fall 2003 and spring 2004 in the Fryingpan River as part of a study to assess the influence of releases from Ruedi Reservoir. Water releases from the impounded reservoir can influence benthic macroinvertebrates and fish communities through regulation of flow and alteration of the thermal regime. It has been hypothesized that erratic changes in discharge have a negative impact on benthic macroinvertebrates (Ptacek et al. 2003); however, it is not clear how the level of discharge during the winter months and the potential formation of anchor ice may influence these communities.

In many ways, the impoundment and physical variables associated with discharge are responsible for the development of an exceptional trout fishery in the Fryingpan River. The purpose of this extended sampling was to evaluate potential impacts associated with low winter flows. This information could be useful when determining management practices that will benefit the trout fishery. Macroinvertebrate monitoring coupled with robust laboratory analysis can provide a great deal of information pertaining to the influence of the flow regime on aquatic conditions.

## **METHODS**

### **MACROINVERTEBRATE SAMPLING**

Benthic macroinvertebrate sampling was conducted during fall (29 October) of 2003, and spring (29 April) of 2004. Two sites on the Fryingpan River (FPR-Res and FPR-TC) were sampled on each occasion. At each location, three samples were taken in riffle habitat using a Hess Sampler with 500  $\mu\text{m}$  mesh to provide quantitative macroinvertebrate data. All samples were taken in areas of similar size substrate and



Figure 1. Satellite image of Fryingspan River study area from Ruedi Reservoir downstream to below Downey Creek (Macroinvertebrates: red, and Thermographs: purple).



Figure 2. Satellite image of Fryingspan River study area from Downey Creek to Basalt (Macroinvertebrates: red, Thermographs: purple).

similar depth to avoid bias that may be directly related to habitat. Depth at each sample location ranged between 24.4 cm and 33.5 cm. Substrate within the Hess Sampler was thoroughly disturbed and individual rocks were scrubbed by hand to dislodge all benthic organisms. Benthic macroinvertebrates were preserved in ethanol and transported to the lab where they were sorted, enumerated and identified to the lowest practical taxonomic level (Merritt and Cummins 1996; Ward et al. 2002).

Identification to the “lowest practical taxonomic level” means that all specimens were identified down to the level that is permitted by the available morphological characteristics. Early life stages of many species lack certain anatomical characteristics that allow the specimen to be identified to the genus or species level. In these cases the “lowest practical taxonomic level” may mean only the family level; however, if the available characteristics are consistent with a species that has been previously confirmed during this study then the individual may be included as a member of that taxa. In these cases the species name is provided in parentheses.

As a means of QA/QC, qualified personnel inspected each sample after sorting and a minimum of 20% of all identified taxa were reviewed. Dr. Boris Kondratieff (Professor of Entomology at Colorado State University) confirmed identifications in all cases where the identification of a specimen was difficult or questionable.

In instances where proper identification was possible, the Orders Ephemeroptera, Plecoptera, and Trichoptera were identified to genus (and many down to the species level). Most specimens of other Orders, including Diptera, were identified to the genus level; however, members of the family Chironomidae were only identified to subfamily or tribe. Further identification would require mounting of head capsules – an expensive and time-consuming process. Data collected were used in various indices recommended by the Rapid Bioassessment Protocols (Plafkin et al. 1989) to provide information regarding macroinvertebrate community structure, function, and general aquatic conditions.



Indices used included Shannon-Weaver diversity (diversity) and evenness (evenness), Family Biotic Index (FBI), EPT index, taxa richness (richness), and description of functional feeding groups. Diversity and evenness values were used to detect changes in macroinvertebrate community structure. In unpolluted waters diversity values typically range from near 3.0 to 4.0. In polluted waters this value is generally less than 1.0. The evenness value ranges between 0.0 and 1.0. Values lower than 0.3 are generally considered indicative of organic pollution (Ward et al. 2002).

The Hilsenhoff Family Biotic Index (FBI) is often used in macroinvertebrate studies as a means of detecting organic enrichment. In this study it was useful for monitoring differences between the sites that may not be attributed to discharge. Because the FBI requires modification for use in many areas, the number indicating a certain water quality rating will vary among regions. Comparison of the values produced within a given system should, however, provide information regarding difference in sites based on nutrient enrichment. Values for the FBI range from 0.0 to 10.0, and increase as water quality decreases (Plafkin et al. 1989).

The Ephemeroptera, Plecoptera, Trichoptera (EPT) index will be employed to assist in the analysis of the data. It is a direct measure of taxa richness among species that are typically considered more sensitive to pollution or other perturbations. This measurement is simply given as the total number of identified taxa in the orders Ephemeroptera, Plecoptera and Trichoptera found at each station.

Taxa richness was also reported for each sampling event during the study. This measurement is reported as the total number of different taxa collected on each date from each sampling location. It is similar to the EPT index, except that it includes all different identifiable benthic macroinvertebrate species. It is useful for describing differences in habitat complexity or aquatic conditions between rivers or site locations.

Benthic macroinvertebrate production at each site was estimated by measuring macroinvertebrate density and biomass. Density was reported as the mean number of

macroinvertebrates/m<sup>2</sup> found at each location. Densities were compared among sites for each sampling occasion. Biomass values were obtained by drying the benthic macroinvertebrates from each sample in an oven at 100° C for 24-hours or until all water content had evaporated. Biomass was reported as the mean dry weight of macroinvertebrates per square meter at each site location. Biomass values provide information in terms of weight of macroinvertebrates produced by habitat at each site. Density and biomass provide a means of measuring and comparing productivity at each sampling location.

Separating invertebrate taxa into functional guilds based on food acquisition provided a measurement of macroinvertebrate community function. Aquatic macroinvertebrates were categorized according to feeding strategy to determine the relative proportion of various groups. The proportion of certain functional feeding groups in the macroinvertebrate community can provide insight to various types of stress in river systems (Ward et al. 2002)

## **THERMAL REGIME**

To describe the winter thermal regime in the Fryingpan River we used Stowaway® Tidbit® temperature loggers (accuracy  $\pm 0.2^{\circ}\text{C}$ ) encased in a small (10 cm) section of pvc pipe for protection. We surveyed water temperatures at the following four locations in the Fryingpan River: downstream of Ruedi Dam at the USGS gaging station (Gaging Station), Pruessing Property (Pruessing Site), Roy Palm Property (Palm Site), and upstream of the confluence with the Roaring Fork River behind Taylor Creek Fly Shop (Fly Shop Site). At each site, capsules were placed in the river and attached to a permanent object by aircraft cable. Holes were drilled in each capsule to ensure adequate circulation of stream water. Each thermograph was set to record hourly water temperatures and was downloaded using a Stowaway® Optic Shuttle. Capsules were placed in inconspicuous mid-channel locations near the stream bottom at a depth where anchor ice is likely to form.

Thermal data was downloaded and input into a computer spreadsheet. We limited the thermal analysis to the months of December, January, and February, which are the months where anchor ice would typically occur. For analysis purposes, we defined an anchor ice event/occurrence as any hourly observation with a water temperature less than 32.3°F (0.2°C).

## **RESULTS**

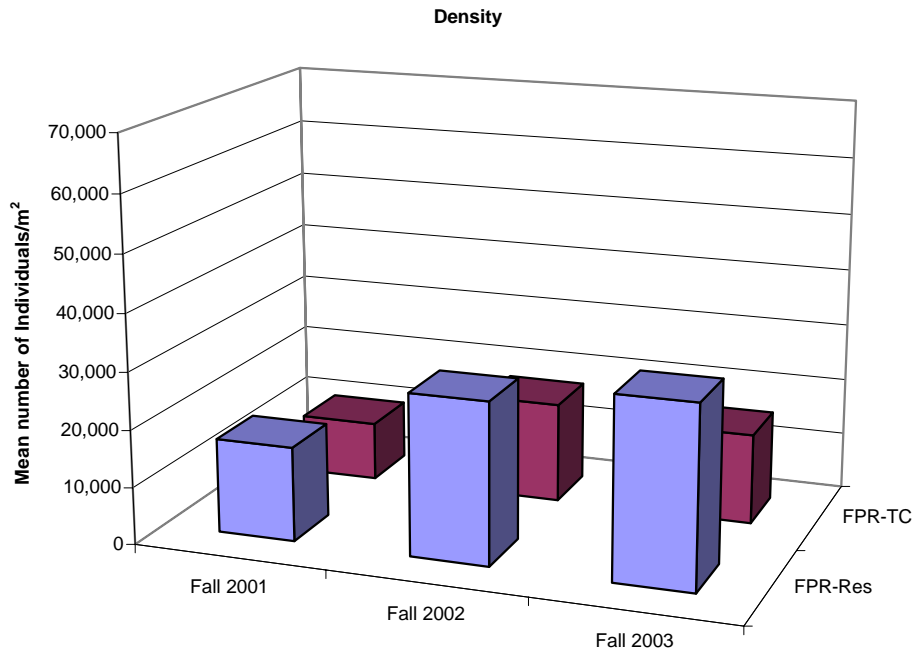
### **FALL 2003**

Macroinvertebrate sampling and analyses was conducted at sites on the Fryingpan River in the fall of 2003 and spring 2004. Population densities and species lists were developed for each sampling site (Appendix A). In general, results of fall 2003 were similar to results from previous years; however, some slight differences were observed (Table 1). Diversity, evenness, and FBI values indicated that conditions had improved slightly in the fall of 2003. There was an increase in density at FPR-RES (Figure 3), and an increase in biomass at FPR-TC (Figure 4). The reason for the inconsistency between these metrics was due to changes in the abundance of specific taxa in each community. The number of small mayflies (*Baetis tricaudatus*) at FPR-RES increased during the fall of 2003, while the density of some of the larger macroinvertebrates declined. This resulted in a slight increase in densities and a slight decrease in biomass at FPR-RES in the fall of 2003. The opposite effect of this process occurred at site FPR-TC. A slight variation in community structure was also reflected in the function analysis (Figure 5). Functional groups exhibited similar composition during all fall sampling events at both sites, with slight variation occurring mostly in the scraper and collector-filterer groups at FPR-TC.

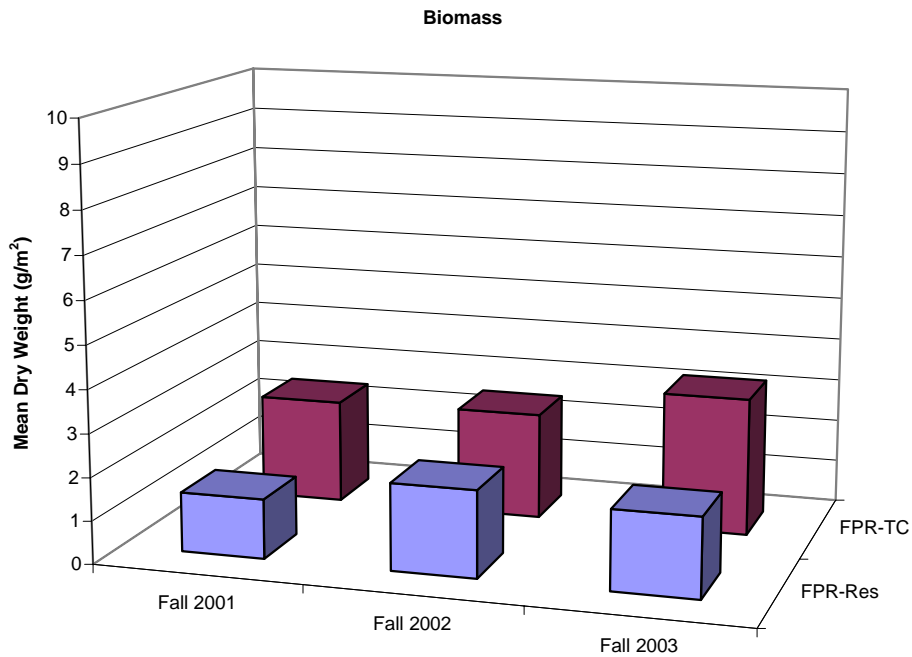
The differences in metrics observed during three years of fall sampling may be well within the range of natural variation that occurs at these sites. Changes in metric values would not be considered substantial, or suggest that a major community altering event had recently occurred (Table 1). It is important to note the yearly similarities among fall samples because it suggests that changes in macroinvertebrate communities in spring samples are the result of events that occur during winter months.

**Table 1. Metrics and comparative values for macroinvertebrate samples collected during the fall season from riffle habitat in the Fryingpan River, Colorado.**

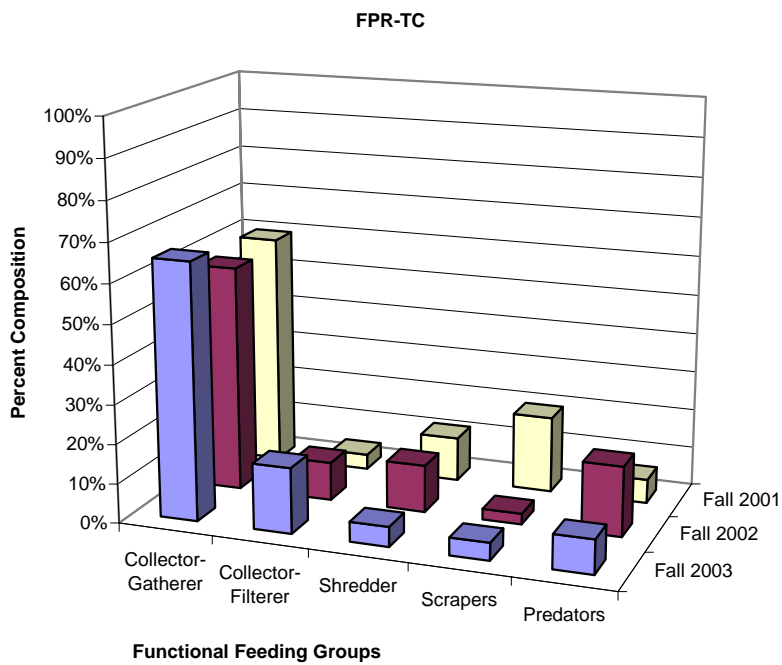
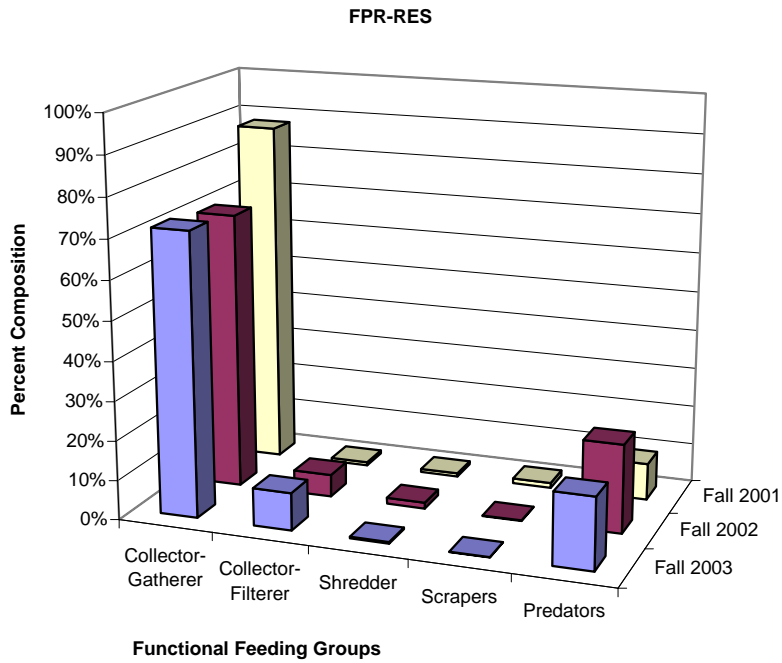
<b>Fall 2001</b>	<b>Diversity</b>	<b>Evenness</b>	<b>FBI</b>	<b>EPT</b>	<b>Taxa Richness</b>	<b>Density (#/m<sup>2</sup>)</b>	<b>Biomass (g/m<sup>2</sup>)</b>
<b>FPR-RES</b>	2.29	0.453	5.86	19	33	16,509	1.3820
<b>FPR-TC</b>	3.76	0.701	4.76	23	41	10,318	2.4338
<b>Fall 2002</b>							
<b>FPR-RES</b>	2.34	0.478	6.62	14	30	28,220	2.0104
<b>FPR-TC</b>	3.35	0.639	5.27	19	38	17,530	2.4856
<b>Fall 2003</b>							
<b>FPR-RES</b>	2.49	0.508	6.35	14	30	31,665	1.8435
<b>FPR-TC</b>	3.39	0.656	5.04	18	36	15,792	3.2179



**Figure 3. Density values obtained from fall sampling at sites on the Fryingpan River, Colorado.**



**Figure 4. Biomass estimates obtained from fall sampling at sites on the Fryingpan River, Colorado.**



**Figure 5. Functional feeding groups at site FPR-RES (top) and FPR-TC (bottom) during fall sampling on the Fryingpan River, Colorado.**

## **SPRING 2004**

Evaluation of data collected during spring 2004 indicated that benthic macroinvertebrate communities at both sites had at least partially recovered from conditions existing in 2003 (Table 2). In general, macroinvertebrate communities exhibited an increase in density and biomass at both sites (Figure 6 and Figure 7), while the changes to community composition were mostly site dependant.

The greatest influence on metric values at FPR- RES during the spring of 2004 resulted from a large increase in the density of mayflies, and the presence of additional EPT taxa (mostly caddisflies) that were not accounted for in the spring of 2003. Results of applied metrics indicated that there was a 31% increase in density and a 33% increase biomass at FPR-RES, while EPT and taxa richness values achieved values that were similar to those reported during 2001 and 2002 (Table 2). Other metrics and the composition of benthic macroinvertebrates based on function have remained relatively consistent at this site (Figure 8). The metrics that remained relatively unaffected (diversity, evenness, FBI and functional feeding groups) are often more sensitive to pollution-related disturbance. These metrics have always indicated some disturbance at FPR-RES that was thought to be an influence of Ruedi Dam.

The applied metrics for site FPR-TC were also influenced by higher densities of macroinvertebrates, but community composition remained similar to that observed in 2003. In the spring of 2004 a 95% gain in density and a 258% increase in biomass were observed at FPR-TC. This resulted from a general increase in abundance of several species, and was not restricted to a specific taxonomic group. The quantity of EPT taxa and individuals in these groups increased at FPR-TC during 2004, but the number of chironomids exhibited a similar trend. Therefore, EPT and taxa richness values increased to levels that would be expected based on the first two years of this study, but diversity, evenness and FBI values remained similar to those reported in 2003 (Table 2). This suggests that the macroinvertebrate community at FPR-TC is still not balanced despite the increase of sensitive taxa. The distribution of functional feeding groups reaffirms

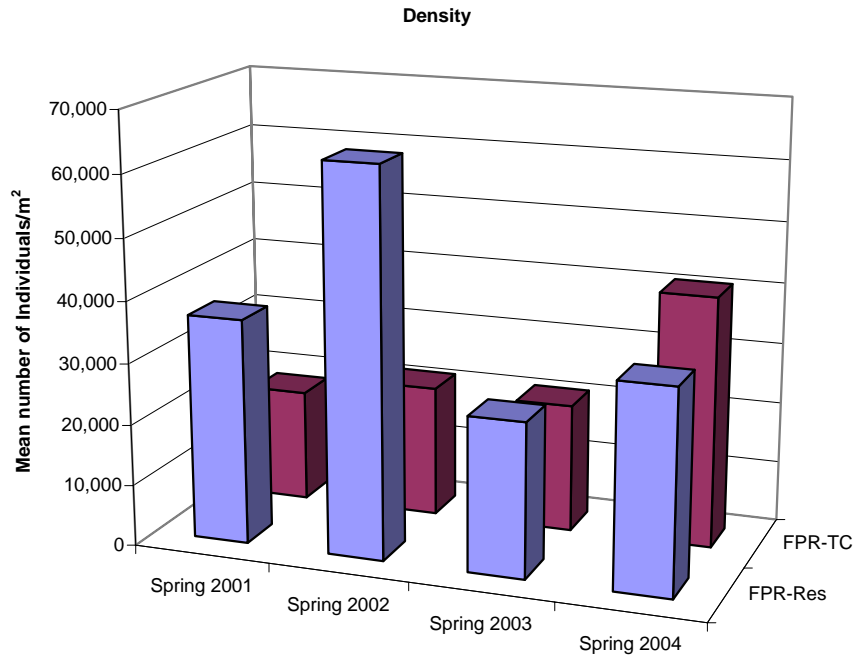
these results by depicting an allocation of species (based on function) that was similar to what was reported in 2003 (Figure 8).

Several of the species that increased in abundance at FPR-TC in the spring 2004 were caddisflies. This is noteworthy because caddisflies are large-bodied insects that contributed substantially to the increase in the mean biomass observed at this site. Caddisflies may be sensitive to anchor ice formation, but are known to be sensitive to rapid changes in discharge. The increase of caddisflies may signify a reduction in rapid flow changes.

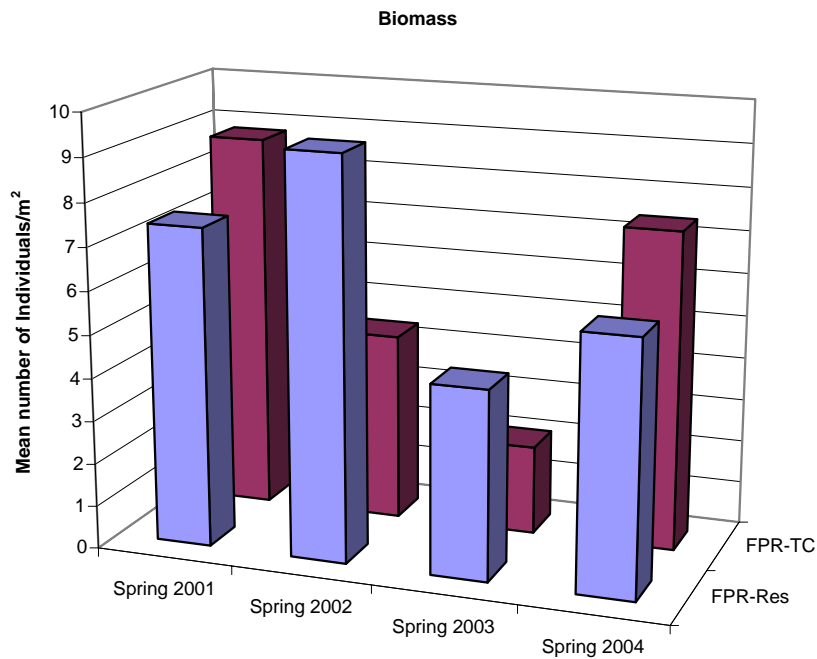


**Table 2. Metrics and comparative values for macroinvertebrate samples collected during the spring season from riffle habitat in the Fryingpan River, Colorado.**

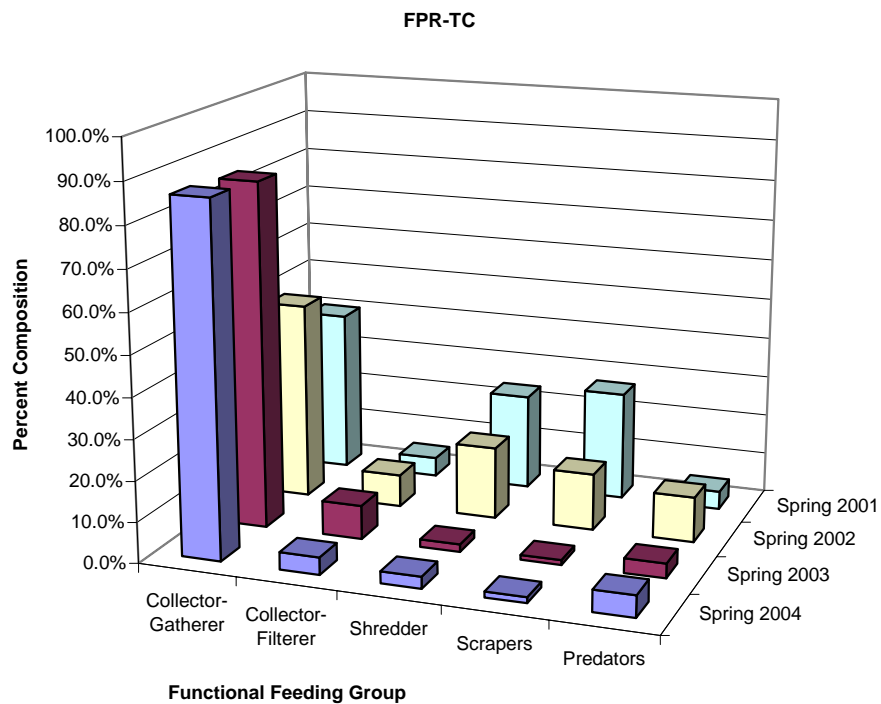
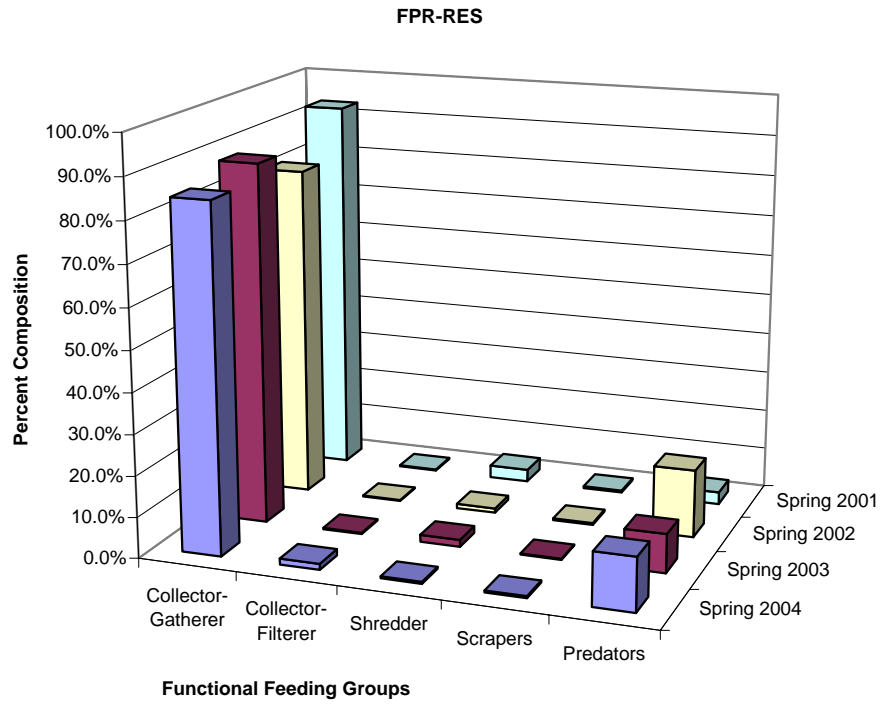
<b>Spring 2001</b>	<b>Diversity</b>	<b>Evenness</b>	<b>FBI</b>	<b>EPT</b>	<b>Taxa Richness</b>	<b>Density (#/m<sup>2</sup>)</b>	<b>Biomass (g/m<sup>2</sup>)</b>
<b>FPR-RES</b>	2.03	0.406	5.72	17	32	36,770	7.4108
<b>FPR-TC</b>	3.71	0.707	3.97	21	38	18,366	8.7948
<b>Spring 2002</b>							
<b>FPR-RES</b>	2.37	0.471	6.06	20	33	62,996	9.2919
<b>FPR-TC</b>	3.66	0.683	4.86	22	41	21,458	4.3774
<b>Spring 2003</b>							
<b>FPR-RES</b>	2.03	0.470	5.90	9	20	25,198	4.3867
<b>FPR-TC</b>	1.93	0.386	5.66	18	32	20,970	2.0629
<b>Spring 2004</b>							
<b>FPR-RES</b>	2.11	0.430	5.50	16	30	33,191	5.8627
<b>FPR-TC</b>	2.11	0.398	5.64	20	39	40,909	7.3951



**Figure 6. Density values obtained from spring sampling at sites on the Fryingpan River, Colorado.**



**Figure 7. Biomass estimates obtained from spring sampling at sites on the Fryingpan River, Colorado.**



**Figure 8. Functional feeding groups at site FPR-RES (top) and FPR-TC (bottom) during spring sampling on the Fryingpan River, Colorado.**

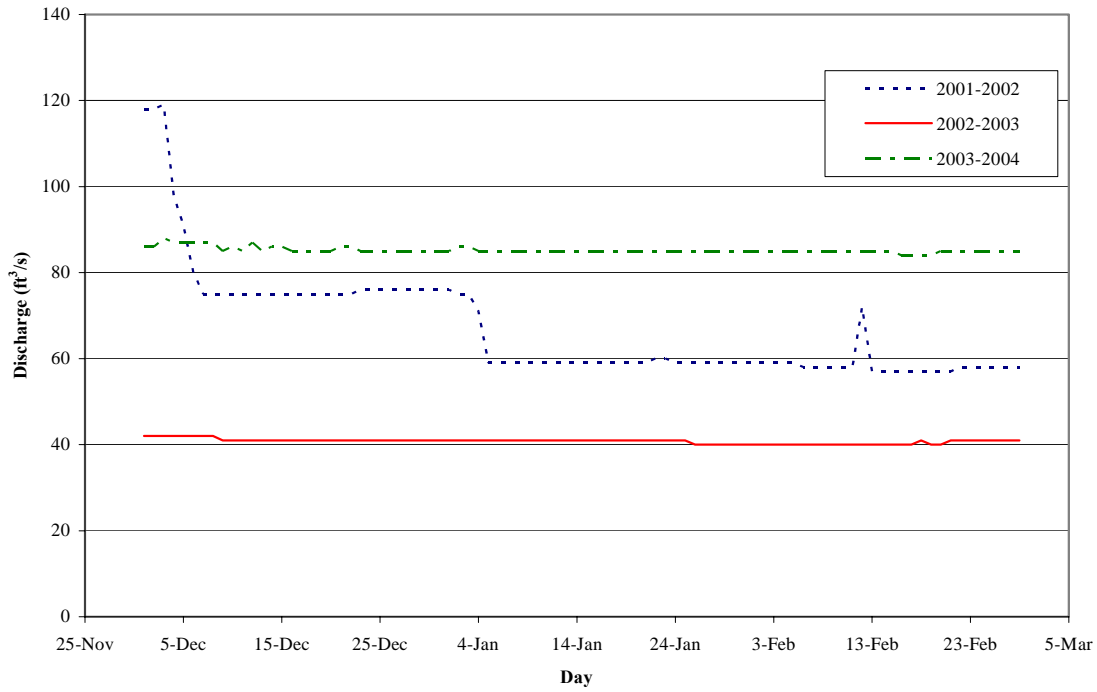
## **DISCUSSION**

Aquatic macroinvertebrate communities were evaluated as a means to elucidate the relationships between winter base flows, anchor ice and macroinvertebrates community structure. The results provide a description of the composition of existing macroinvertebrate communities at the time and location of sampling. The mechanisms that influence the community assemblages are numerous and include variables not directly related to flow manipulations (biological interactions, air temperature, etc.). However, the direct and indirect effects of the flow regime resulting from the regulated discharge in the Fryingpan River appear to influence benthic macroinvertebrate communities.

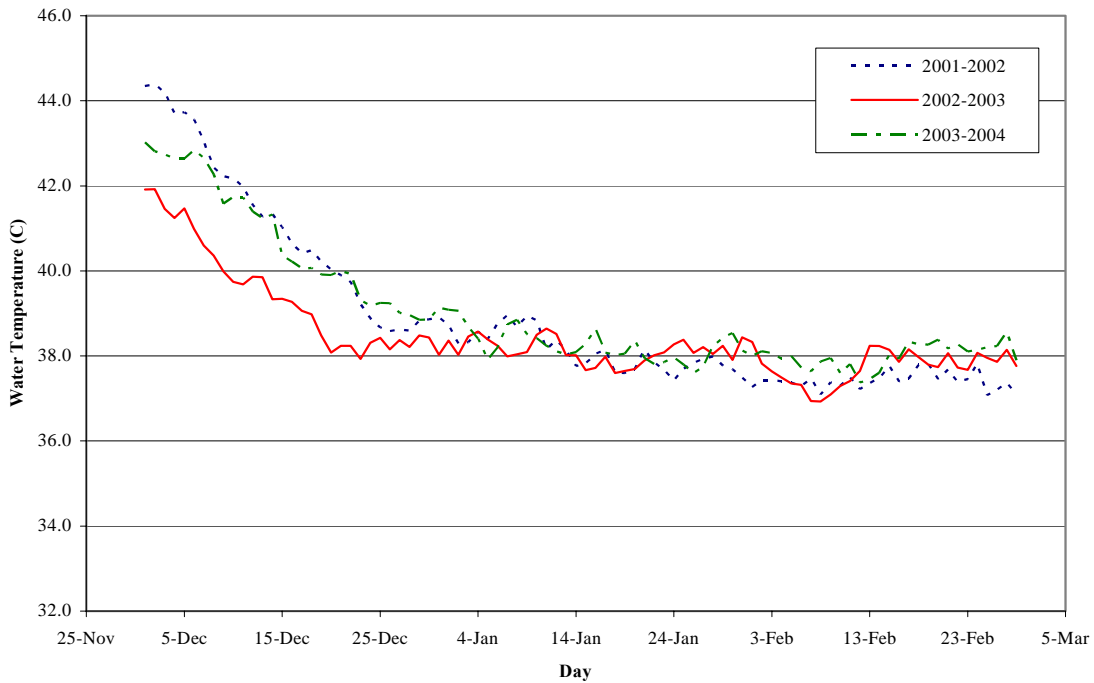
The magnitude of discharge may be the most important factor that influences macroinvertebrates during the winter months. In the winter of 2002-2003 base flows were recorded at an average 40.8 cfs below Ruedi Dam from December through February (Figure 9). Metrics used to describe benthic macroinvertebrate communities in the spring indicated that conditions had declined at both sites. It was hypothesized that benthic communities in the spring of 2003 were responding to physical processes associated with lower discharge (Rees et al. 2003). During the winter of 2003-2004 the mean discharge was approximately 85.2 cfs and some apparent recovery of macroinvertebrate communities was observed at both sites on the Fryingpan River.

Results of metric values from site FPR-RES are likely influenced primarily by discharge because water temperature does not allow anchor ice formation at this site. The mean daily water temperature below the dam was slightly lower during the early portion of the 2002-2003 winter, but water temperature during the coldest months has been similar during each winter season of this study (Figure 10).

Although macroinvertebrate impact and recovery seem to be associated with the magnitude of discharge at both sites on the Fryingpan River, the data suggests that the community at FPR-TC is also influenced by some indirect effects of discharge.



**Figure 9. Winter discharge (December-February) for the Fryingpan River below Ruedi Reservoir, Colorado.**



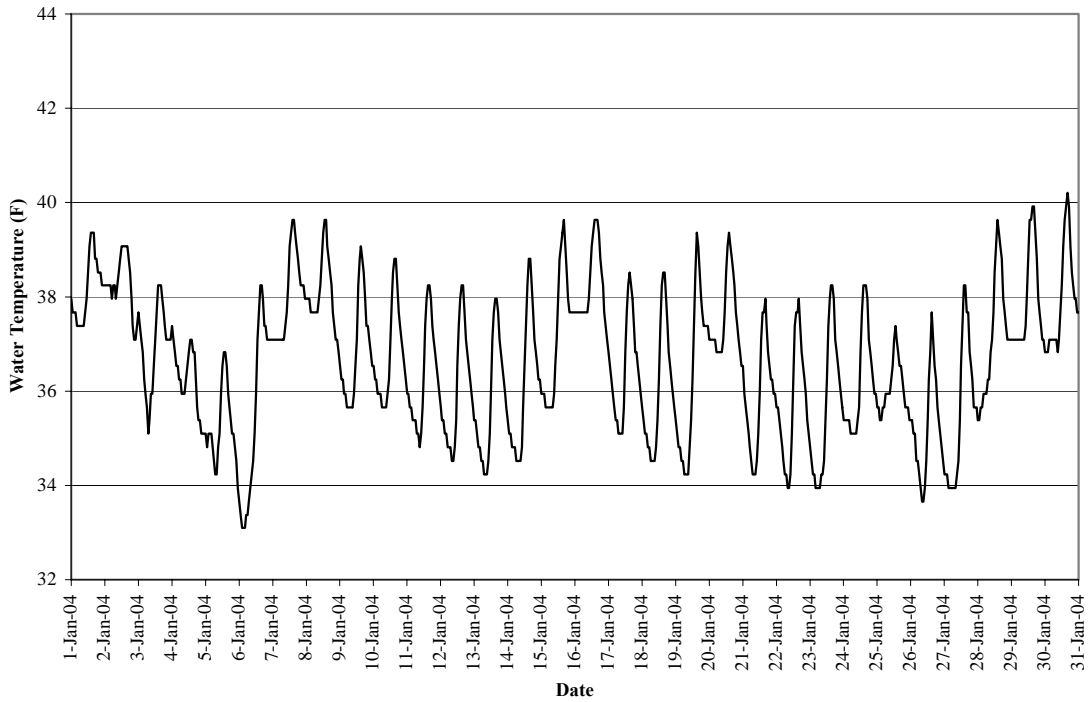
**Figure 10. Winter water temperatures for Fryingpan River below Ruedi Reservoir, Colorado.**

The formation and frequency of occurrence of anchor ice at FPR-TC appears to be a contributing influence on macroinvertebrate community structure and function.

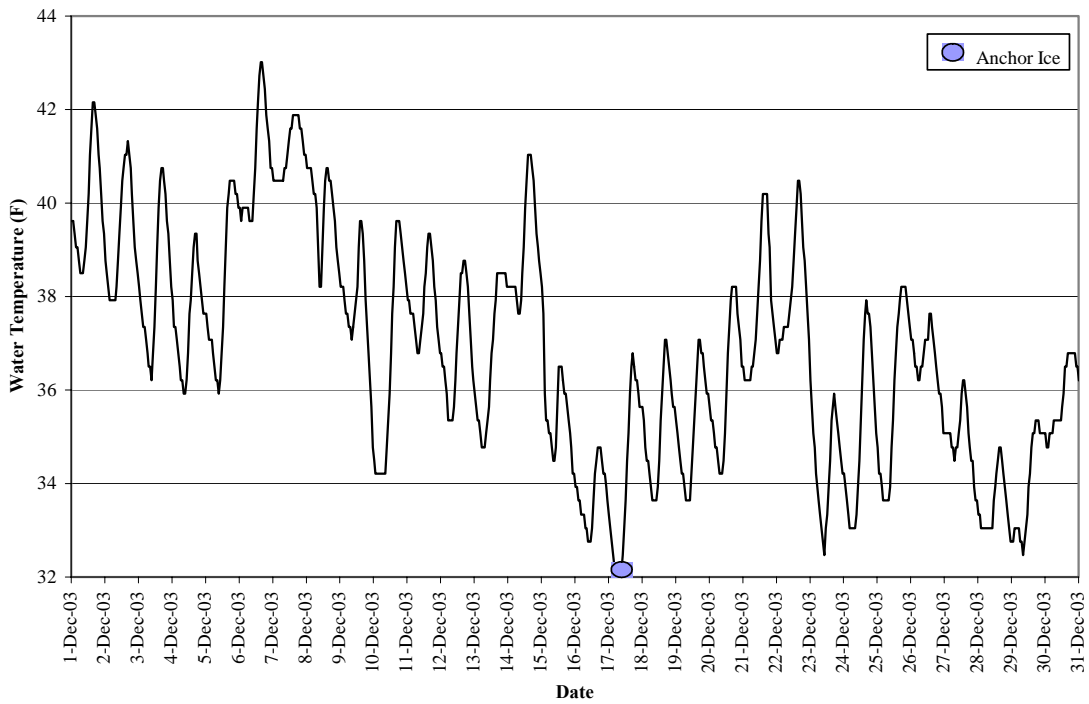
The responses of metric values to the low flows in the winter of 2002-2003 were noticeably different between sites. At FPR-TC densities of EPT taxa were reduced while chironomid densities increased. This was thought to be a result of combined affects from low flows and anchor ice formation (Rees et al. 2003). Decreases observed in the density of EPT taxa may have been the result of a scouring process that resulted from anchor ice formation. Many of the EPT taxa use microhabitat (surface areas) that would be susceptible to this scouring process, whereas the habitat used by most chironomids (subsurface) provides better protection.

The results of sampling in 2004 after higher winter flows indicated that densities of many EPT taxa had recovered but chironomid numbers had increased as well. This recent data suggests that two or more concurrent winters with higher flows may be necessary to achieve an optimum balance in the macroinvertebrate community at FPR-TC.

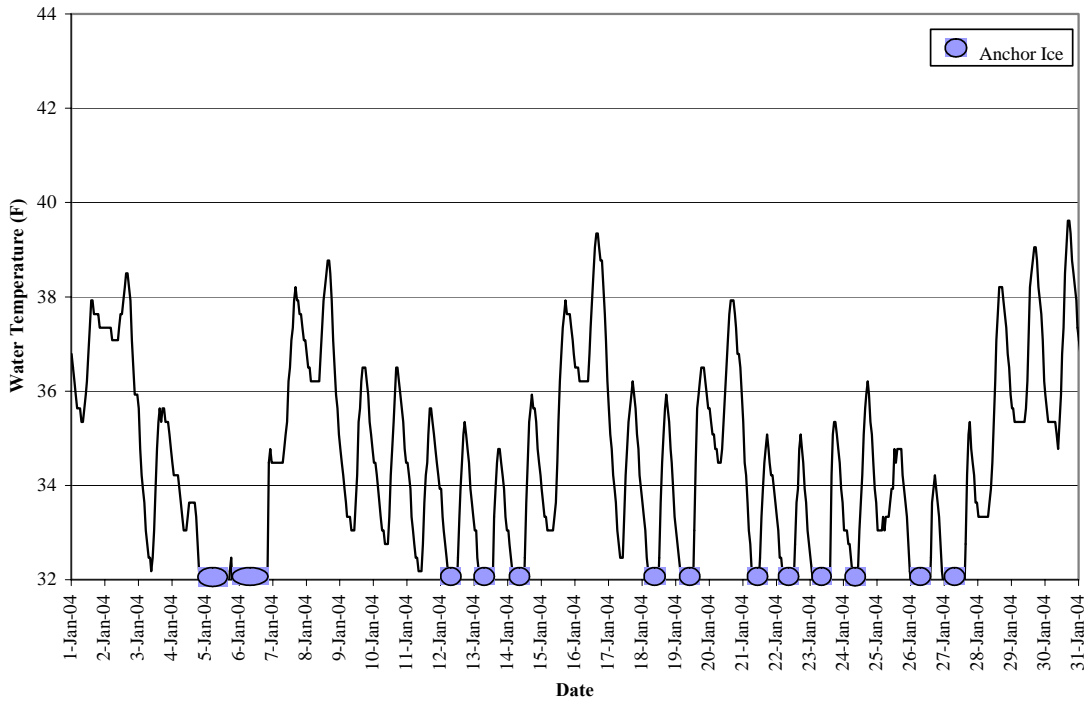
Because anchor ice is known to have a negative impact on aquatic biota it is important to identify causes and areas of potential formation. Thermograph data from January 2004 (the coldest month of that winter) at the Pruessing Site (located between the dam and Taylor Creek confluence) suggested that anchor ice formation did not occur between the dam and this location at a discharge of 85 cfs (Figure 11). Thermograph data from December 2003, January 2004, and February 2004 identified periods of anchor ice formation immediately upstream of the FPR-TC site (Figures 12-14). Thermograph data from the Fryingpan River in Basalt indicated an increased frequency and duration of anchor ice formation (Figures 15-17). The frequency of occurrence and duration of anchor ice formation seems to increase with distance downstream.



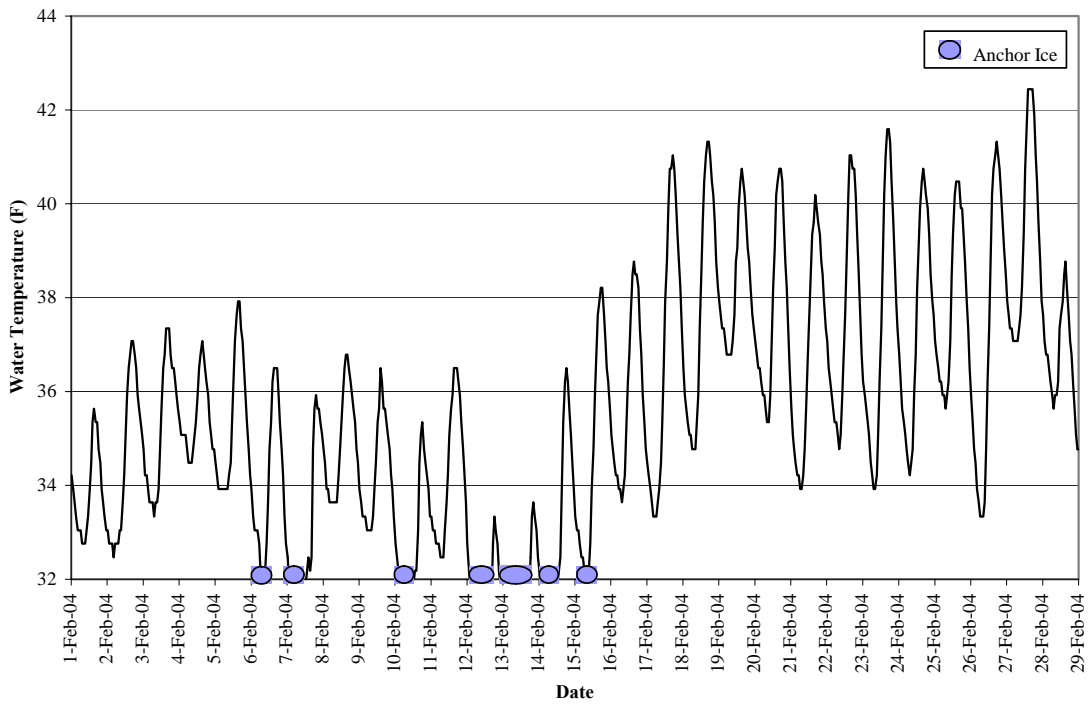
**Figure 11. Hourly water temperatures during January 2004 on the Fryingpan River, Pruessing Site.**



**Figure 12. Hourly water temperatures during December 2003 at Palm Site.**

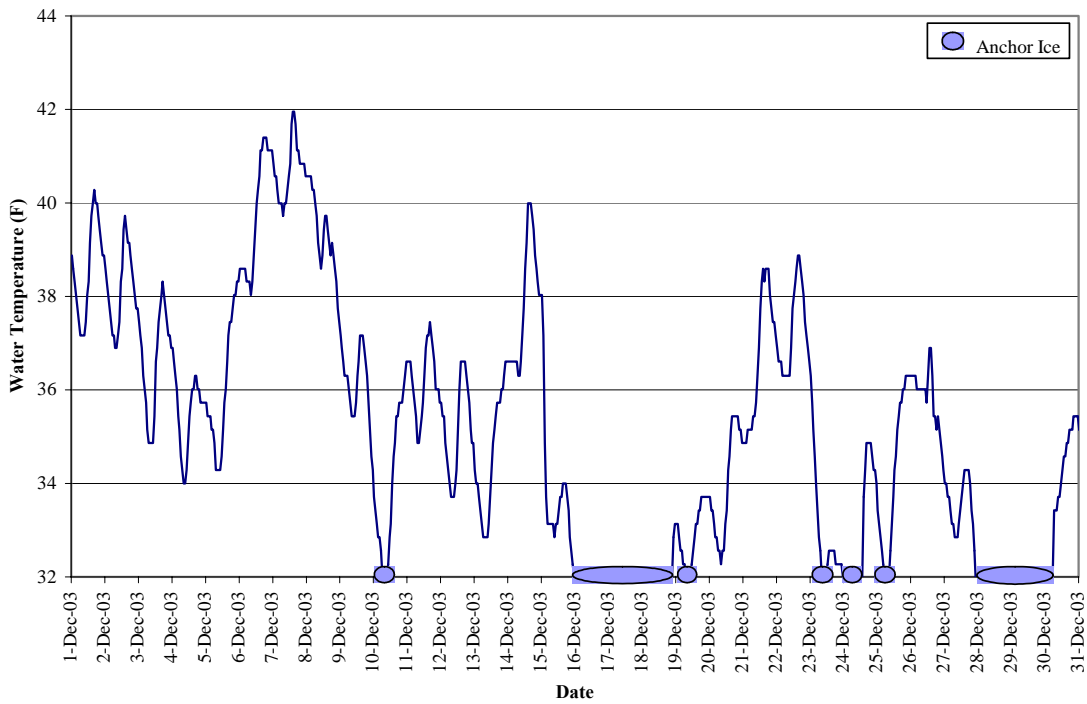


**Figure 13. Hourly water temperatures during January 2004 at Palm Site.**

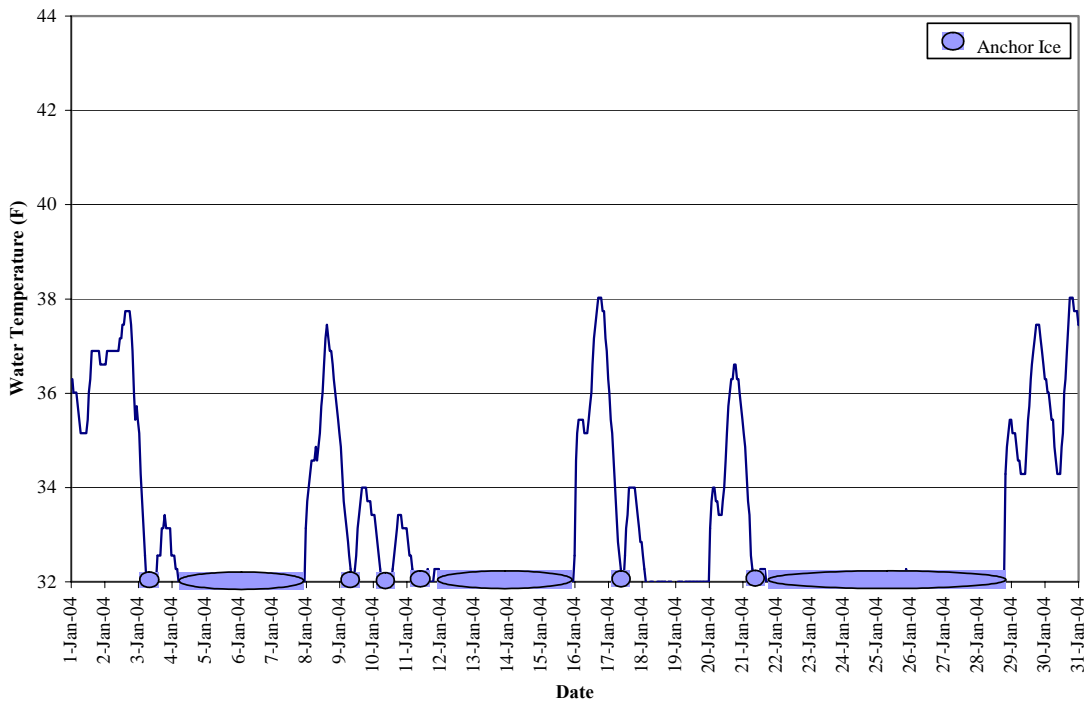


**Figure 14. Hourly water temperatures during February 2004 at Palm Site.**

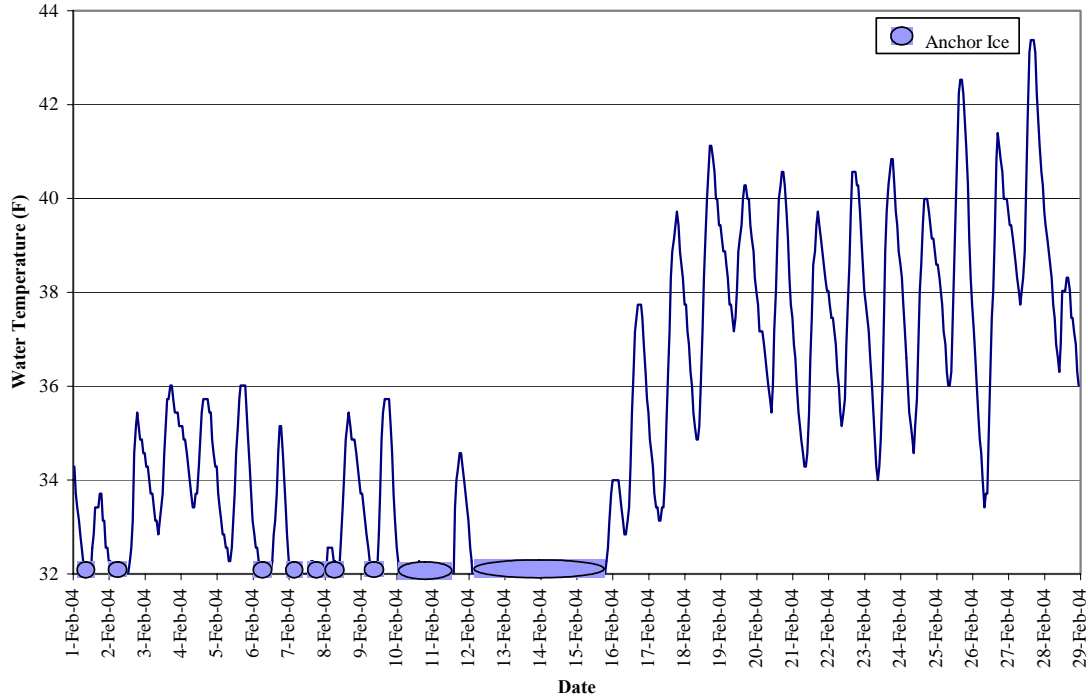




**Figure 15. Hourly water temperatures during December 2003 on the Fryingspan River, at Basalt.**



**Figure 16. Hourly water temperatures during January 2004 on the Fryingspan River, at Basalt.**



**Figure 17. Hourly water temperatures during February 2004 on the Fryingpan River, at Basalt.**

Results of this study suggest that magnitude of discharge and air temperature work together to influence anchor ice formation. Thermograph data from two consecutive winter seasons at the Palm site (immediately upstream of FPR-TC) indicated that anchor ice formation at this location was much less frequent during the winter of 2003-2004 compared to the previous winter (Table 3). The average length of an anchor ice occurrence was also much less in 2003-2004. In 2003-2004, each event averaged 10.7 hours compared to 25.7 hours during the 2002-2003 winter. It is possible that the magnitude of the effect of anchor ice formation on the macroinvertebrate community may be amplified as the length of the event increases.

The available data suggests that air temperatures were similar in the study area during both winters (Table 4), but discharge was quite different (Figure 9). The lower discharge at site FPR-TC in 2002-2003 was much more conducive to the formation of anchor ice than the higher flows during the following winter. This results in the location where

anchor ice first appears to be further upstream in years with lower flows than those years with higher winter flows. In addition to an increase the length of river susceptible to anchor ice formation, the magnitude of the event also increases with low flows.

The available data suggest that anchor ice was at least partially responsible for the degraded condition of the macroinvertebrate community at FPR-TC during the spring of 2003. To alleviate anchor ice related stress to the macroinvertebrate community, an effort should be made to avoid low wintertime releases out of Ruedi Reservoir.

**Table 3. Number of anchor ice occurrences (hourly water temperature less than 0.2°C) during winters (December-February) of 2002-2003 and 2003-2004 at Palm Site.**

Month	Year	
	2002-2003	2003-2004
December	229	6
January	214	164
February	200	86

**Table 4. Average monthly air temperature (°F) recorded at Aspen, Colorado (Station: Aspen 1 SW, Coop ID: 050372).**

Month/Year	Average Max Temperature (°F)	Average Min Temperature (°F)	Count of days with Min Temperature ≤ 5°F
December 01	35.2	8.2	13
January 02	35	8.6	10
February 02	37	5.2	12
December 02	36.5	12.6	6
January 03	40.8	16.3	0
February 03	36.8	12.3	5
December 03	36.6	12.3	7
January 04	36.5	8.8	8
February 04	37.4	10.2	8

## RECOMMENDATIONS

Questions for future studies include:

- What are the impacts to benthic macroinvertebrates in lower reaches of the Fryingpan River (near Basalt)?
- There is a need to further evaluate the specific relationship between anchor ice formation, discharge and air temperature at site FPR-TC (or other locations in the Fryingpan River)?
- In a winter with average air temperatures and cloud cover, what is the discharge that would be necessary to maintain high densities of macroinvertebrates.
- What is the time required for recovery that can be expected after impact to macroinvertebrate communities?

To answer the preceding questions, macroinvertebrate sampling and air and water temperature monitoring should continue in the spring and fall for at least the next year. Ideally, it would be good to compare sampling before and after winters with different base lows and different air temperature patterns. Sites for temperature monitoring and macroinvertebrate sampling should at least include FPR-TC, and could be expanded to the lower river. Water quality data (collected frequently) from this reach of the Fryingpan River would also be useful.

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## **ACKNOWLEDGEMENTS**

We would like to thank the project partners, the Roaring Fork Conservancy, Colorado River Water Conservation District, and Ruedi Water and Power Authority for their continued vision, and the support necessary to conduct this study on the Fryingpan River. In addition, we are grateful to the Bureau of Reclamation for helping to fund this phase of the study. Finally, we would like to thank individuals and landowners that assisted with river access, particularly Roy Palm, Bart Chandler and Tim Heng.

## **APPENDIX A**

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**Table 1. Macroinvertebrate data collected from the Fryingpan River at site FPR-RES on 29 October 2003.**

Fryingpan River										
FPR-RES		Sample		Mean	ni*LOGni	Count				FBI
29 Oct. 03	1	2	3							
<i>Acentrella insignificans</i>	2	3	2	2.33	0.86	1	4			0.0034
<i>Baetis (flavistriga)</i>										
<i>Baetis (tricaudatus)</i>	641	600	501	580.67	1604.92	1	4			0.8441
<i>Drunella grandis</i>	1	1	2	1.33	0.17	1	1			0.0005
<i>Drunella coloradensis</i>										
<i>Drunella doddsi</i>		4	1	1.67	0.37	1	1			0.0006
<i>Ephemerella sp.</i>	10	12		7.33	6.35	1	1			0.0027
<i>Cinygmula sp.</i>										
<i>Epeorus longimanus</i>	1			0.33	-0.16	1	4			0.0005
<i>Rhithrogena sp.</i>										
<i>Paraleptophlebia sp.</i>	1	1	1	1.00	0.00	1	2			0.0007
<i>Tricorythodes minutus</i>	2	1		1.00	0.00	1	4			0.0015
<i>Caenis sp.</i>										
<i>Pteronarcella badia</i>										
<i>Capnia sp.</i>										
<i>Zapada sp.</i>	3			1.00	0.00	1	2			0.0007
<i>Paraperla frontalis</i>										
<i>Sweltsa sp.</i>										
<i>Triznaka signata</i>										
<i>Claassenia sabulosa</i>										
<i>Hesperoperla pacifica</i>										
<i>Skwala americana</i>										
<i>Isoperla fulva</i>										
<i>Isoperla sp. 2</i>										
<i>Brachycentrus americanus</i>	2			0.67	-0.12	1	1			0.0002
<i>Brachycentrus occidentalis</i>										
<i>Agapetus boulderensis</i>										
<i>Culoptila sp.</i>										
<i>Glossosoma sp.</i>										
<i>Arctopsyche grandis</i>			1	0.33	-0.16	1	4			0.0005
<i>Hydropsyche cockerelli</i>										
<i>Hydropsyche occidentalis</i>										
<i>Hydropsyche sp. (oslari)</i>										
<i>Hydroptila sp.</i>	13	2	6	7.00	5.92	1	1			0.0025
<i>Ochrotrichia sp.</i>										
<i>Lepidostoma sp.</i>	1			0.33	-0.16	1	4			0.0005
<i>Ceraclea sp.</i>										
<i>Oecetis sp.</i>										
<i>Dolophilodes aequalis</i>										
<i>Rhyacophila brunnea</i>										
<i>Rhyacophila coloradensis</i>		1	1	0.67	-0.12	1	0			0.0000
<i>Neothremma alicia</i>										
<i>Oligophlebodes minuta</i>										
Orthoclaadiinae	698	1278	997	991.00	2969.11	1	6			2.1609
Tanypodinae										
Tanytarsini	31	39	17	29.00	42.41	1	6			0.0632
Chironomini										
Diamesinae	54	19	16	29.67	43.68	1	6			0.0647
<i>Simulium sp.</i>	400	175	112	229.00	540.40	1	6			0.4993
<i>Protanyderus margarita</i>										
<i>Chelifera sp.</i>										
<i>Clinocera sp.</i>		1	1	0.67	-0.12	1	6			0.0015
<i>Hemerodromia sp.</i>										
<i>Antocha sp.</i>	13	35	23	23.67	32.52	1	3			0.0258
<i>Dicranota sp.</i>										
<i>Hexatoma sp.</i>										
<i>Tipula sp.</i>	1			0.33	-0.16	1	3			0.0004
<i>Atherix pachypus</i>										
<i>Pericoma sp.</i>										
<i>Optioservus sp.</i>		1		0.33	-0.16	1	4			0.0005
<i>Heterimnius corpulentus</i>	13	8	7	9.33	9.05	1	4			0.0136
<i>Zaitzevia parvula</i>										
<i>Microcyloepus sp.</i>										
<i>Narpus concolor</i>										
<i>Hydracarina sp.</i>	7	8	4	6.33	5.08	1	8			0.0184
<i>Gammarus sp.</i>	4	3	3	3.33	1.74	1	4			0.0048
<i>Physa sp.</i>			1	0.33	-0.16	1	8			0.0010
<i>Pisidium sp.</i>	4	2	4	3.33	1.74	1	8			0.0097
<i>Dugesia sp.</i>										
<i>Polycelis coronata</i>	296	659	499	484.67	1301.54	1	8			1.4091
<i>Oligochaeta</i>	201	567	222	330.00	831.11	1	10			1.1993
<i>Nematoda</i>	6	3	6	5.00	3.49	1	10			0.0182
Totals	2405.0	3423.0	2427.0	2751.67	7399.16	30				6.35
Shannon Weaver Diversity					2.49					
Shannon Weaver Evenness					0.508					



**Table 3. Macroinvertebrate data collected from the Fryingpan River at site FPR-RES on 29 April 2004.**

Fryingpan River									
FPR-RES		Sample		Mean	ni*LOGni	Count			FBI
29 Apr. 04	1	2	3						
<i>Acentrella insignificans</i>	5			1.67	0.37	1	4		0.0023
<i>Baetis (flavistriga)</i>									
<i>Baetis (tricaudatus)</i>	843	1044	1462	1116.33	3402.35	1	4		1.5481
<i>Drunella grandis</i>		2	1	1.00	0.00	1	1		0.0003
<i>Drunella coloradensis</i>			1	0.33	-0.16	1	1		0.0001
<i>Drunella doddsi</i>	2	4	1	2.33	0.86	1	1		0.0008
<i>Ephemerella sp.</i>	9	14	14	12.33	13.46	1	1		0.0043
<i>Serratella tibialis</i>									
<i>Cinygmula sp.</i>	16	9	9	11.33	11.95	1	4		0.0157
<i>Epeorus longimanus</i>	8	8	6	7.33	6.35	1	4		0.0102
<i>Rhithrogena sp.</i>									
<i>Paraleptophlebia sp.</i>	1	1	4	2.00	0.60	1	2		0.0014
<i>Tricorythodes minutus</i>									
<i>Pteronarcella badia</i>									
<i>Prostoia besametsa</i>									
<i>Zapada sp.</i>	1			0.33	-0.16	1	2		0.0002
<i>Triznaka signata</i>									
<i>Sweltsa sp.</i>									
<i>Claassenia sabulosa</i>									
<i>Hesperoperla pacifica</i>									
<i>Isoperla fulva</i>									
<i>Isoperla sp. 2</i>									
<i>Skwala americana</i>									
<i>Brachycentrus americanus</i>		1		0.33	-0.16	1	1		0.0001
<i>Brachycentrus occidentalis</i>									
<i>Micrasema bactro</i>									
<i>Culoptila sp.</i>									
<i>Glossosoma sp.</i>		2	1	1.00	0.00	1	0		0.0000
<i>Arctopsyche grandis</i>									
<i>Hydropsyche cockerelli</i>									
<i>Hydropsyche occidentalis</i>									
<i>Hydropsyche sp. (oslari)</i>									
<i>Hydroptila sp.</i>		1	1	0.67	-0.12	1	1		0.0002
<i>Lepidostoma sp.</i>			1	0.33	-0.16	1	4		0.0005
<i>Ceraclea sp.</i>									
<i>Oecetis sp.</i>									
<i>Rhyacophila brunnea</i>		2	3	1.67	0.37	1	0		0.0000
<i>Rhyacophila coloradensis</i>			2	0.67	-0.12	1	0		0.0000
<i>Neothremma alicia</i>									
<i>Oligophlebodes minuta</i>									
Orthoclaidiinae	512	764	1832	1036.00	3123.91	1	6		2.1551
Tanypodinae									
Tanytarsini		3		1.00	0.00	1	6		0.0021
Chironomini			1	0.33	-0.16	1	8		0.0009
Diamasinae	90	232	312	211.33	491.34	1	6		0.4396
<i>Simulium sp.</i>	23	55	48	42.00	68.18	1	6		0.0874
<i>Chelifera sp.</i>									
<i>Clinocera sp.</i>									
<i>Hemerodromia sp.</i>									
<i>Oreogeton sp.</i>									
<i>Tipula sp.</i>									
<i>Antocha sp.</i>	1	7	11	6.33	5.08	1	3		0.0066
<i>Dicranota sp.</i>									
<i>Hexatoma sp.</i>									
<i>Atherix pachypus</i>									
<i>Pericoma sp.</i>									
<i>Optioservus sp.</i>		1		0.33	-0.16	1	4		0.0005
<i>Heterolimnius corpulentus</i>	2	2	2	2.00	0.60	1	4		0.0028
<i>Zaitzevia parvula</i>									
<i>Narpus concolor</i>									
<i>Hydracarina sp.</i>	2	8	11	7.00	5.92	1	8		0.0194
<i>Gammarus sp.</i>		1	1	0.67	-0.12	1	4		0.0009
<i>Physa sp.</i>									
Planorbidae									
<i>Pisidium sp.</i>	1			0.33	-0.16	1	8		0.0009
<i>Dugesia sp.</i>									
<i>Polycelis coronata</i>	477	351	257	361.67	925.25	1	8		1.0031
Oligochaeta	37	71	51	53.00	91.39	1	10		0.1838
Nematoda		4	4	2.67	1.14	1	10		0.0092
Totals	2030.0	2587.0	4036.0	2884.33	8147.64	30			5.50
Shannon Weaver Diversity					2.11				
Shannon Weaver Evenness					0.430				

**Table 4. Macroinvertebrate data collected from the Fryingpan River at site FPR-TC on 29 April 2004.**

Fryingpan River										
FPR-TC		Sample			Mean	ni*LOGni	Count			FBI
29 Apr. 04	1	2	3							
<i>Acentrella insignificans</i>										
<i>Baetis (flavistriga)</i>	22	15	16	17.67	22.03	1	4			0.0199
<i>Baetis (tricaudatus)</i>	307	179	479	321.67	806.55	1	4			0.3619
<i>Drunella grandis</i>	9	5	23	12.33	13.46	1	1			0.0035
<i>Drunella coloradensis</i>										
<i>Drunella doddsi</i>										
<i>Ephemera sp.</i>	16	13	27	18.67	23.73	1	1			0.0053
<i>Serratella tibialis</i>	33	12	35	26.67	38.03	1	1			0.0075
<i>Cinygmula sp.</i>	35	19	7	20.33	26.60	1	4			0.0229
<i>Epeorus longimanus</i>	15	14	8	12.33	13.46	1	4			0.0139
<i>Rhithrogena sp.</i>										
<i>Paraleptophlebia sp.</i>	69	27	65	53.67	92.83	1	2			0.0302
<i>Tricorythodes minutus</i>										
<i>Pteronarcella badia</i>										
<i>Prostoia besametsa</i>		1		0.33	-0.16	1	2			0.0002
<i>Zapada sp.</i>										
<i>Triznaka signata</i>										
<i>Sweltsa sp.</i>										
<i>Claassenia sabulosa</i>										
<i>Hesperoperla pacifica</i>	3	2	2	2.33	0.86	1	1			0.0007
<i>Isoperla fulva</i>		1	2	1.00	0.00	1	2			0.0006
<i>Isoperla sp. 2</i>										
<i>Skwala americana</i>										
<i>Brachycentrus americanus</i>	48	33	78	53.00	91.39	1	1			0.0149
<i>Brachycentrus occidentalis</i>										
<i>Micrasema bactro</i>										
<i>Culoptila sp.</i>										
<i>Glossosoma sp.</i>		4		1.33	0.17	1	0			0.0000
<i>Arctopsyche grandis</i>	10	12	22	14.67	17.11	1	4			0.0165
<i>Hydropsyche cockerelli</i>	1			0.33	-0.16	1	4			0.0004
<i>Hydropsyche occidentalis</i>										
<i>Hydropsyche sp. (oslari)</i>		1		0.33	-0.16	1	4			0.0004
<i>Hydroptila sp.</i>										
<i>Lepidostoma sp.</i>	128	37	90	85.00	164.00	1	4			0.0956
<i>Ceraclea sp.</i>										
<i>Oecetis sp.</i>										
<i>Rhyacophila brunnea</i>	6	1	4	3.67	2.07	1	0			0.0000
<i>Rhyacophila coloradensis</i>			1	0.33	-0.16	1	0			0.0000
<i>Neothremma alicia</i>										
<i>Oligophlebodes minuta</i>	15	10	11	12.00	12.95	1	4			0.0135
<i>Orthocladiinae</i>	2209	2378	2666	2417.67	8179.92	1	6			4.0805
<i>Tanypodinae</i>	39	9	22	23.33	31.92	1	6			0.0394
<i>Tanytarsini</i>	71	10	27	36.00	56.03	1	6			0.0608
<i>Chironomini</i>		11		3.67	2.07	1	8			0.0083
<i>Diamasinae</i>	2	1	4	2.33	0.86	1	6			0.0039
<i>Simulium sp.</i>	1	1		0.67	-0.12	1	6			0.0011
<i>Chelifera sp.</i>	7		1	2.67	1.14	1	6			0.0045
<i>Clinocera sp.</i>										
<i>Hemerodromia sp.</i>										
<i>Oreogeton sp.</i>			3	1.00	0.00	1	6			0.0017
<i>Tipula sp.</i>										
<i>Antocha sp.</i>	24	49	86	53.00	91.39	1	3			0.0447
<i>Dicranota sp.</i>	1		1	0.67	-0.12	1	3			0.0006
<i>Hexatoma sp.</i>										
<i>Atherix pachypus</i>		3		1.00	0.00	1	2			0.0006
<i>Pericoma sp.</i>		1		0.33	-0.16	1	10			0.0009
<i>Optioservus sp.</i>	3	4	5	4.00	2.41	1	4			0.0045
<i>Heterlimnius corpulentus</i>	86	41	44	57.00	100.08	1	4			0.0641
<i>Zaitzevia parvula</i>										
<i>Narpus concolor</i>										
<i>Hydracarina sp.</i>	7	8	19	11.33	11.95	1	8			0.0255
<i>Gammarus sp.</i>										
<i>Physa sp.</i>										
<i>Planorbidae</i>										
<i>Pisidium sp.</i>	68	32	36	45.33	75.09	1	8			0.1020
<i>Dugesia sp.</i>										
<i>Polycelis coronata</i>	184	80	157	140.33	301.32	1	8			0.3158
<i>Oligochaeta</i>	99	98	91	96.00	190.30	1	10			0.2700
<i>Nematoda</i>	2		1	1.00	0.00	1	10			0.0028
<b>Totals</b>	<b>3520.0</b>	<b>3112.0</b>	<b>4033.0</b>	<b>3555.00</b>	<b>10368.65</b>	<b>39</b>				<b>5.64</b>
Shannon Weaver Diversity					2.11					
Shannon Weaver Evenness					0.399					