



**Supplemental Report**

**A Study of the Ecological Processes  
on the Fryingpan and Roaring Fork Rivers  
Related to Operation of Ruedi Reservoir**

**Prepared for**

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## **INTRODUCTION**

The mechanisms that govern the structure and function of benthic macroinvertebrate communities in the Fryingpan River are not entirely understood. For this reason, sampling continued during the Fall 2002 and Spring 2003 as a supplement to the study that occurred during the previous seasons (Ptacek et al. 2003). Releases from the impoundment at Ruedi Reservoir directly influence the benthic macroinvertebrate and fish communities that exist downstream. The influence of regulated discharge on the macroinvertebrate community can result in an increase in thermal stability and extended periods of flow stability. However, it is unknown how the level of discharge during the winter months and the potential formation of anchor ice may influence these communities.

In many ways, the impoundment and physical variables associated with discharge are responsible for the development of an exceptional trout fishery in the Fryingpan River. The purpose of this extended sampling was to evaluate potential impacts associated with low winter flows. This information could be useful when determining management practices that will benefit the trout fishery. The results provided by macroinvertebrate monitoring and accurate identifications can provide a great deal of information pertaining to the flow regime and other aquatic conditions.

## **METHODS**

### **MACROINVERTEBRATE SAMPLING**

Benthic macroinvertebrate sampling was conducted during fall (12 Oct.) of 2002, and spring (29 Apr.) of 2003. Two sites on the Fryingpan River (FPR-Res and FPR-TC) were sampled on each occasion. At each location three samples were taken in riffle habitat using a Hess Sampler with 500  $\mu\text{m}$  mesh to provide quantitative macroinvertebrate data. All samples were taken in areas of similar size substrate and similar depth to avoid bias that may be directly related to habitat. Depth at each sample location ranged between 24.4 cm and 33.5 cm. Substrate within the Hess Sampler was



Figure 1. Satellite image of Fryingpan River study area from Ruedi Reservoir downstream to below Downey Creek (Macroinvertebrates: red, Thermographs: purple).



Figure 2. Satellite image of Fryingpan River study area from Downey Creek to Basalt (Macroinvertebrates: red, Thermographs: purple).

thoroughly disturbed and individual rocks were scrubbed by hand to dislodge all benthic organisms. Benthic macroinvertebrates were preserved in ethanol and transported to the lab where they were sorted, enumerated and identified to the lowest practical taxonomic level (Merritt and Cummins 1996; Ward et al. 2002).

Identification to the “lowest practical taxonomic level” means that all specimens were identified down to the level that is permitted by the available morphological characteristics. Early life stages of many species sometimes lack certain anatomical characteristics that allow the specimen to be identified to the genus or species level. In these cases the “lowest practical taxonomic level” may mean only the family level; however, if the available characteristics are consistent with a species that has been previously confirmed during this study then the individual may be included as a member of that taxa. In these cases the species name is provided in parentheses.

As a means of QA/QC, qualified personnel inspected each sample after sorting and a minimum of 20% of all identified taxa were reviewed. Dr. Boris Kondratieff (Professor of Entomology at Colorado State University) confirmed identifications in all cases where the identification of a specimen was difficult or questionable.

In instances where proper identification was possible, the Orders Ephemeroptera, Plecoptera, and Trichoptera were identified to genus (and many down to the species level). Most specimens of other Orders, including Diptera, were identified to the genus level; however, members of the family Chironomidae were only identified to subfamily or tribe. Further identification would require mounting of head capsules – an expensive and time-consuming process. Data collected were used in various indices recommended by the Rapid Bioassessment Protocols (Plafkin et al. 1989) to provide information regarding macroinvertebrate community structure, function, and general aquatic conditions. Population densities and species lists were developed for each sampling site. A description of the metrics used in this study is provided in the final report.

## **THERMAL REGIME**

To describe the thermal regime in the Fryingpan River we used Stowaway® Tidbit® temperature loggers (accuracy  $\pm 0.2^{\circ}\text{C}$ ) encased in a small (10 cm) section of pvc pipe for protection. Capsules were placed in the river and attached to a permanent object by aircraft cable. Each capsule was drilled with holes to ensure adequate water circulation. Each thermograph was set to record hourly water temperature and was downloaded every few months using a Stowaway® Optic Shuttle™. Capsules were placed in out-of-the-way and inconspicuous mid-channel locations near the stream bottom at a depth where anchor ice is likely to form.

## **RESULTS**

Additional macroinvertebrate sampling and analyses were conducted at sites on the Fryingpan River in the fall of 2002 and spring 2003 (Tables 1 and 2). Species lists are provided in Appendix A. The following section describes the changes observed in the applied metrics.

### **FALL 2002**

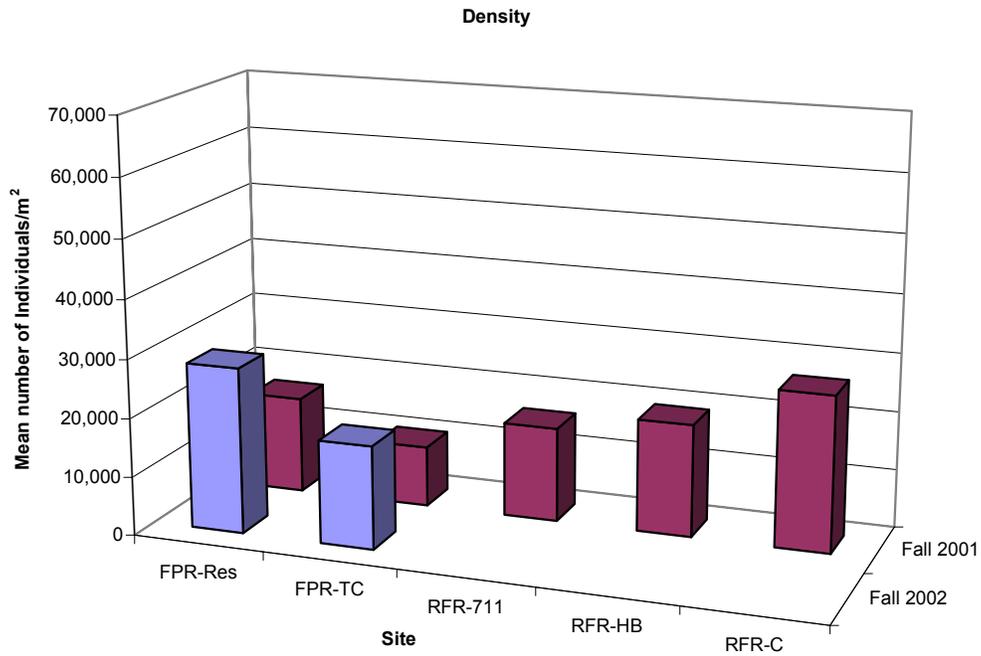
In general, results of fall 2002 were similar to results the previous year; however, some slight differences were observed in taxa richness, density, and biomass (Table 1). Results of the fall 2002 sampling suggested that taxa richness had decreased slightly at both sights while density and biomass had increased. An increase in density (Figure 3) and biomass (Figure 4) at the FPR-RES site was particularly evident. The increase in these metrics can mostly be attributed to an increase in the number of flatworms (*P. coronata*) and chironomids that were collected during the fall of 2002. This slight variation in community structure was also reflected in community function (Figure 5). Functional groups exhibited similar composition during both years with slight variation occurring mostly in the scraper and predator groups.

**Table 1. Metrics and comparative values for macroinvertebrate samples collected during the fall season from riffle habitat in the Fryingpan River, Colorado.**

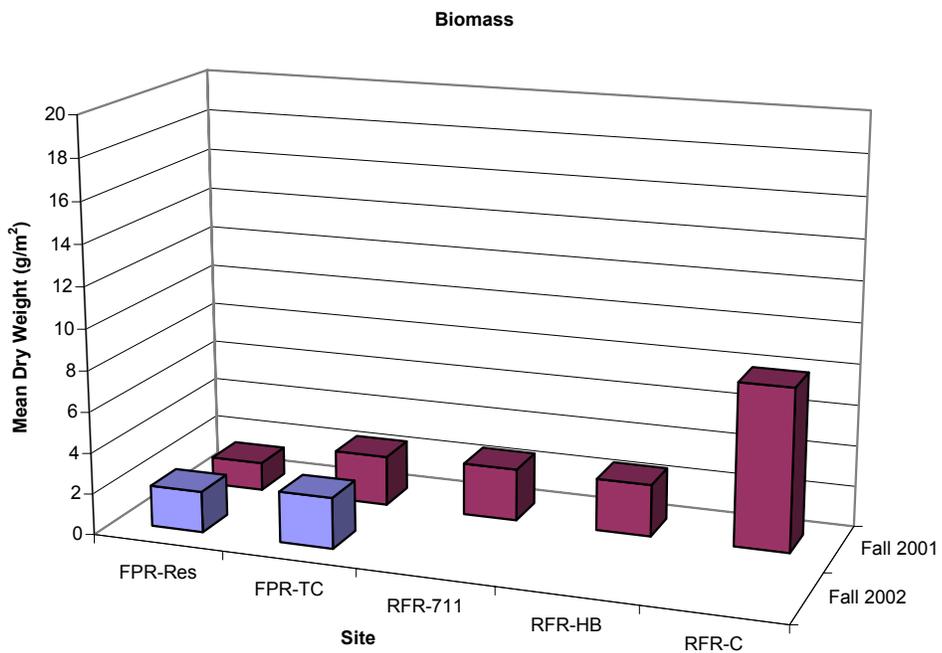
Fall 2001	Diversity	Evenness	FBI	EPT	Taxa Richness	Density (#/m <sup>2</sup> )	Biomass (g/m <sup>2</sup> )
FPR-RES	2.29	0.453	5.86	19	33	16,509	1.3820
FPR-TC	3.76	0.701	4.76	23	41	10,318	2.4338
<b>Fall 2002</b>							
FPR-RES	2.34	0.478	6.62	14	30	28,220	2.0104
FPR-TC	3.35	0.639	5.27	19	38	17,530	2.4856

**Table 2. Metrics and comparative values for macroinvertebrate samples collected during the spring season from riffle habitat in the Fryingpan River, Colorado.**

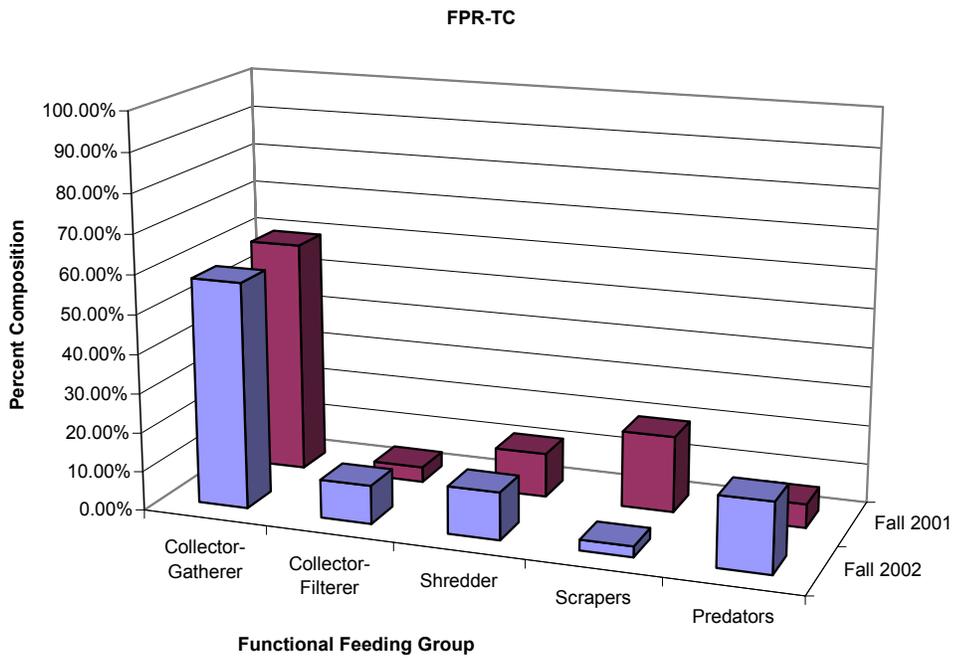
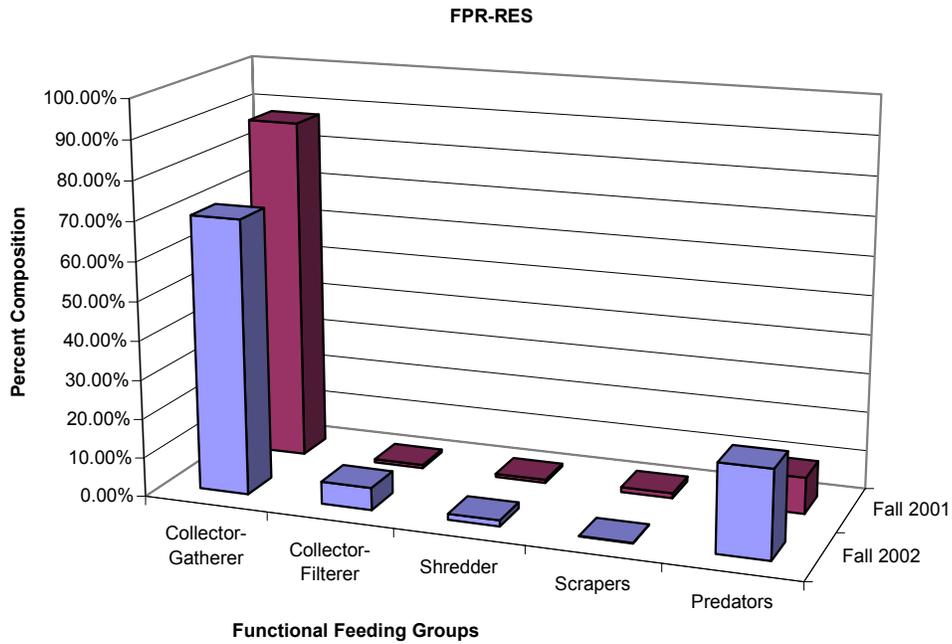
Spring 2001	Diversity	Evenness	FBI	EPT	Taxa Richness	Density (#/m <sup>2</sup> )	Biomass (g/m <sup>2</sup> )
FPR-RES	2.03	0.406	5.72	17	32	36,770	7.4108
FPR-TC	3.71	0.707	3.97	21	38	18,366	8.7948
<b>Spring 2002</b>							
FPR-RES	2.37	0.471	6.06	20	33	62,996	9.2919
FPR-TC	3.66	0.683	4.86	22	41	21,458	4.3774
<b>Spring 2003</b>							
FPR-RES	2.03	0.470	5.90	9	20	25,198	4.3867
FPR-TC	1.93	0.386	5.66	18	32	20,970	2.0629



**Figure 3. Density values obtained from fall sampling at sites on the Fryingpan River, Colorado.**



**Figure 4. Biomass estimates obtained from fall sampling at sites on the Fryingpan River, Colorado.**



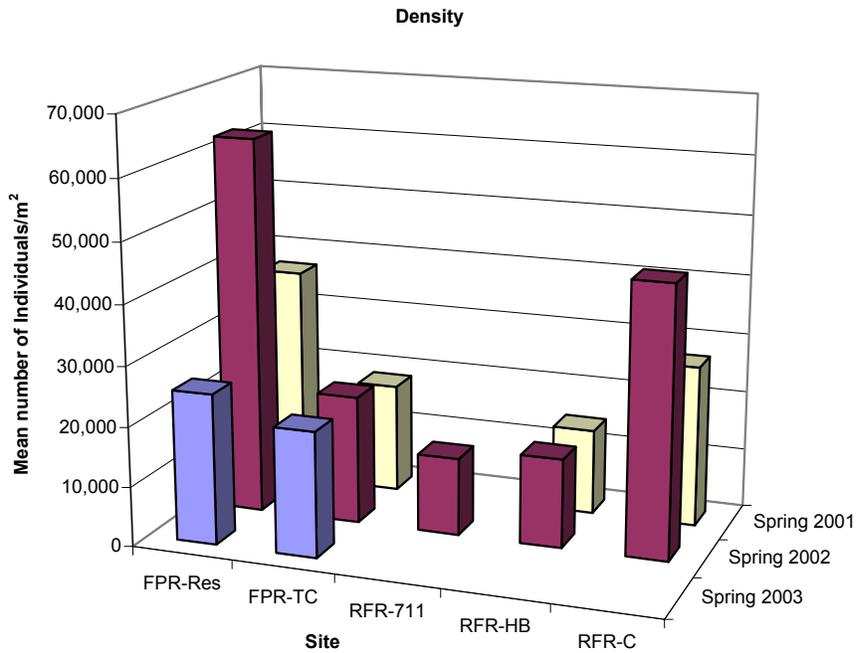
**Figure 5. Functional feeding groups at site FPR-RES (top) and FPR-TC (bottom) during fall sampling on the Fryingpan River, Colorado.**

The differences in metrics observed during two years of fall sampling may be well within the range of natural variation that occurs at these sites. It is difficult to determine what would be considered “normal”, or what may cause slight variation among years using data from only two years. It is important to note that fall samples were similar between 2001 and 2002 when observing the substantial changes that occurred in spring samples (Table 1).

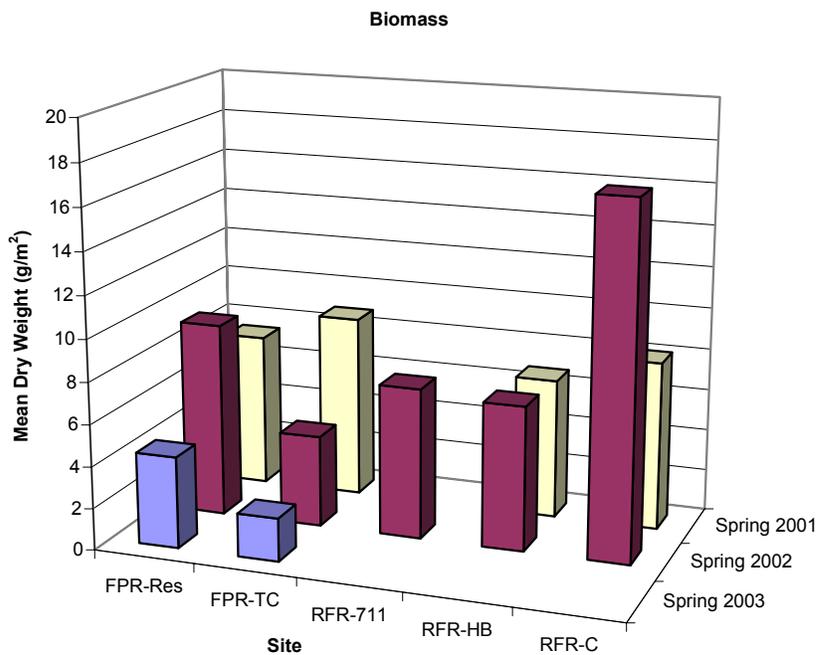
### **SPRING 2003**

Evaluation of data collected during spring 2003 indicated that there had been substantial changes in macroinvertebrate communities compared to results from the previous spring sampling events (Table 2). The lack of similarities in response of the metric values at each site suggests that the changes in benthic macroinvertebrate communities were somewhat different at each location.

Some index values indicated substantial change in benthic macroinvertebrate communities at site FPR-RES (EPT, taxa richness, density and biomass), while others did not (diversity, evenness, FBI and functional feeding groups). The EPT and taxa richness indices were the result of the lack of stoneflies and caddisflies that were found in samples during spring 2003. These two Orders of aquatic insects accounted for eight (8) of the taxa collected in 2001 and ten (10) of the taxa collected in 2002. Only one (1) representative from these two Orders was collected in 2003. Density of benthic macroinvertebrates at FPR-RES decreased by more than 60% from what had been reported the previous year (Figure 6). The low density combined with the absence of larger size macroinvertebrates (stoneflies and caddisflies) resulted in a biomass number (4.3867 g) that was considerably lower than those reported for the previous spring samples (Figure 7). The metrics that remained relatively unaffected (diversity, evenness, FBI and functional feeding groups) are often more sensitive to pollution-related disturbance. These metrics have always indicated some disturbance that was thought to be an influence of the Ruedi Dam.



**Figure 6. Density values obtained from spring sampling at sites on the Fryingpan River, Colorado.**

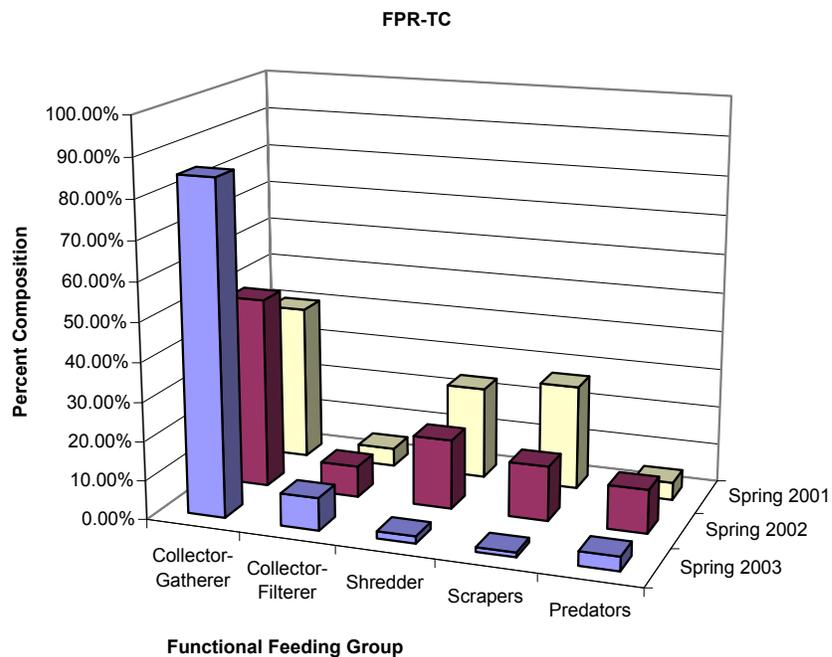
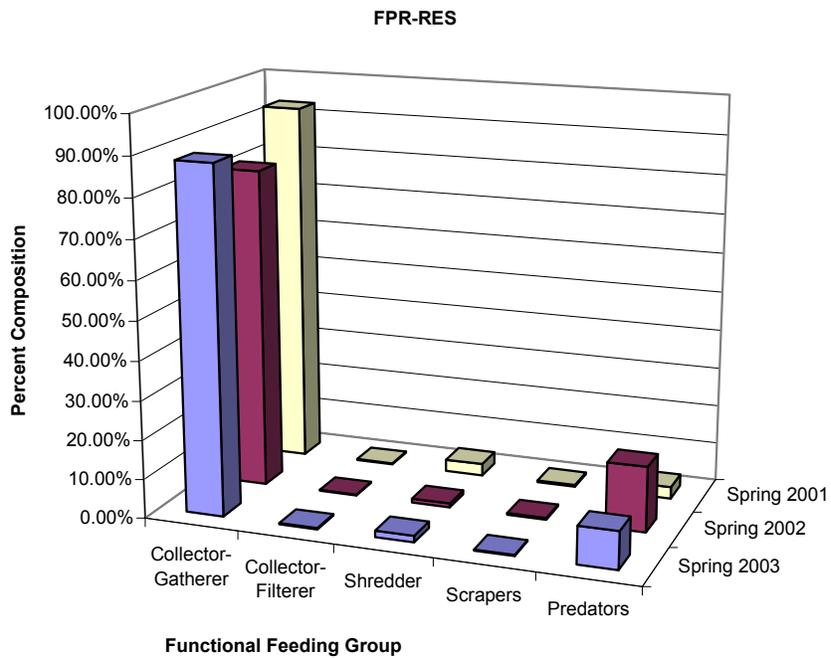


**Figure 7. Biomass estimates obtained from spring sampling at sites on the Fryingpan River, Colorado.**

The data provided by spring sampling during 2003 at site FPR-TC indicated a decline in aquatic conditions that was reflected by most metric values (Table 2). This change in metric values was the result of a severe reduction in the density of EPT taxa (mayflies, stoneflies and caddisflies) in spring samples during 2003. The data from the appendix in this and the previous report shows that the average number of EPT individuals collected in a sample during 2001 was 1,144. This number declined slightly in the spring of 2002 to 1,052. In the spring of 2003 the average number of individuals representing the EPT taxa had decreased to 261. At the same time as the density of EPT taxa was decreasing, the density of chironomids was increasing. This resulted in a mean density value that was similar to those reported in previous years (Figure 6), but the replacement of large-bodied taxa (stoneflies, caddisflies) with small-bodied taxa (chironomids) was resulted in a decrease in the mean biomass (Figure 7). The disproportion of chironomids to other taxa caused a decrease in diversity and evenness values (1.93 and 0.386, respectively) to levels that indicate a stressed or impacted community. Ward et al. (2002) indicates that optimum metric values range between 3.0 - 4.0 (for diversity), and 0.6 – 0.8 (evenness) in Colorado streams. The greater proportion of chironomids resulted in a shift in the composition of functional feeding guilds to a community dominated by collector-gatherers (Figure 8).

## **DISCUSSION**

Aquatic macroinvertebrate communities were evaluated as a means to understand the relationships between winter base flows, anchor ice and macroinvertebrates community structure. The results provide a description of the composition of existing macroinvertebrate communities at the time and location of sampling. The mechanisms that influence the community assemblages are numerous and include variables not directly related to flow manipulations (biological interactions, air temperature, etc.). However, the direct and indirect effects of the flow regime and regulated discharge in the Fryingpan River provide a major influence on benthic macroinvertebrates.

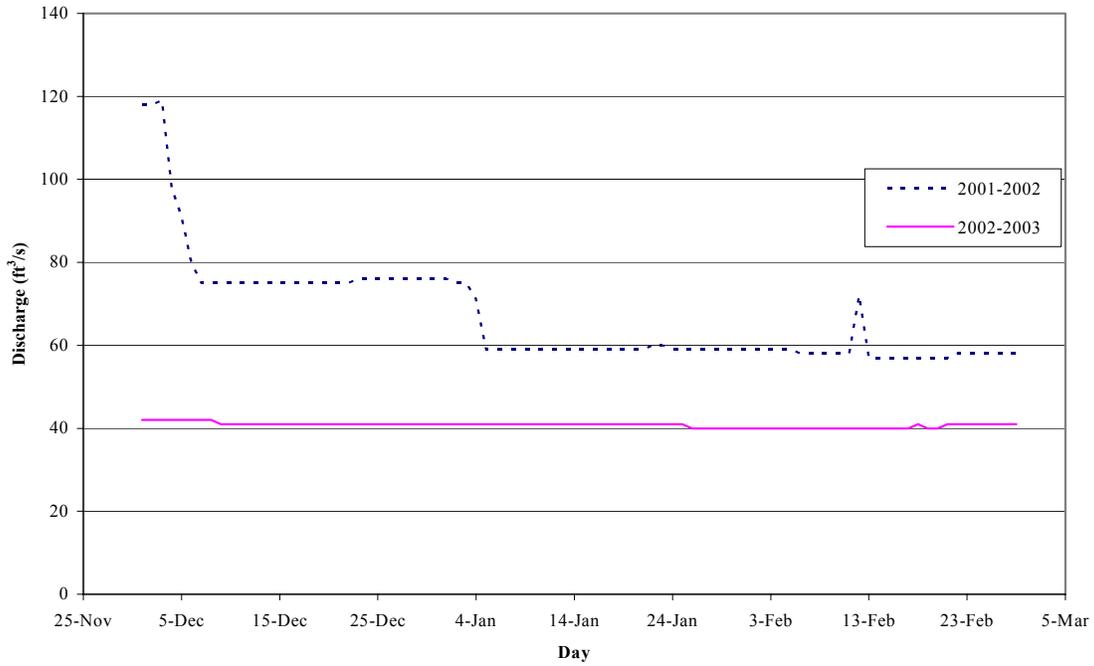


**Figure 8. Functional feeding groups at site FPR-RES (top) and FPR-TC (bottom) during spring sampling on the Fryingpan River, Colorado.**

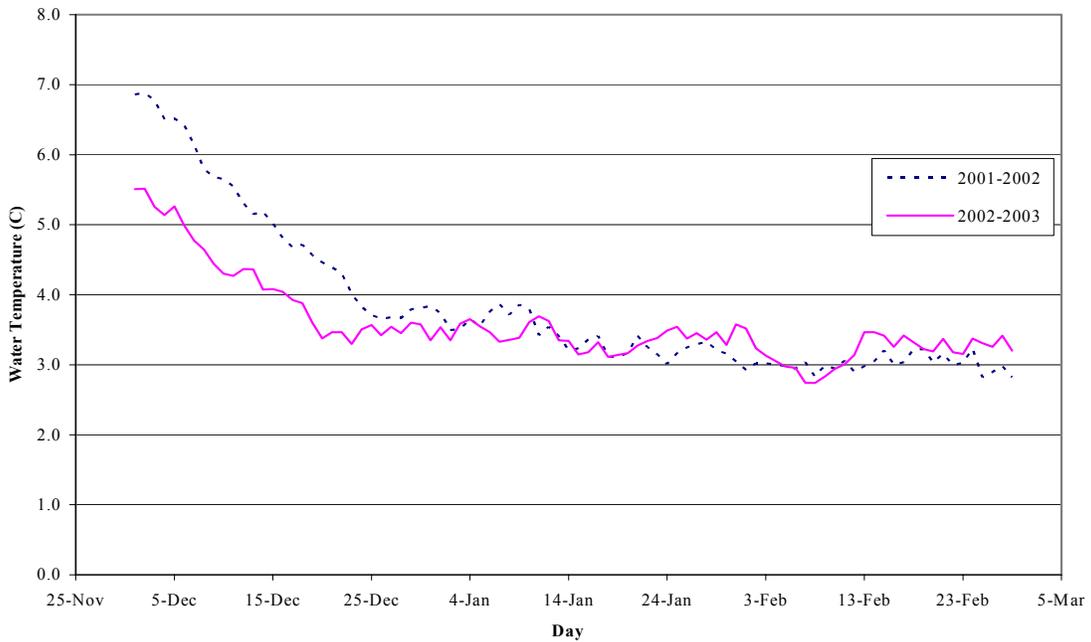
Changes that were observed in benthic macroinvertebrate communities during the spring of 2003 were similar to those expected from various types of pollution. However, the lack of similar responses between sites FPR-RES and FPR-TC, and the season in which the degrading of these communities took place, did not support the idea that that these alterations were pollution induced. This suggests that benthic communities in the spring of 2003 were responding to physical processes associated with changes in discharge.

Discharge during the winter of 2002-2003 was unusually low due to drought conditions during previous years, (Figure 9). The difference in discharge between the winter 2001/2002 and 2002/2003 seasons may be directly responsible for much of the observed alterations in benthic macroinvertebrate communities. At site FPR-RES there are few other explanations for a decrease in sensitive taxa and a substantial decline in density. The mean daily water temperature below the dam was slightly lower during the early portion of the 2002-2003 winter (Figure 10), but anchor ice at this location should not have been an issue. Some other possible explanations could include changes in water quality (which was not measured) or human influence (increased wading traffic from fishermen).

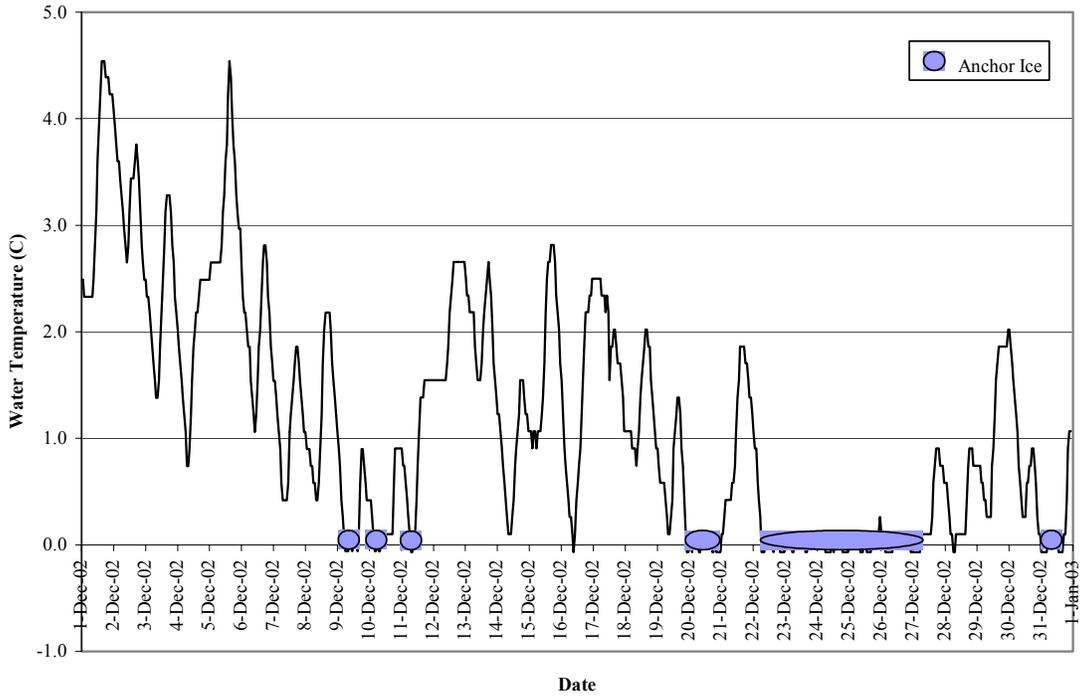
The impact to benthic macroinvertebrate communities at site FPR-TC was different than that observed at FPR-RES. Densities of EPT taxa were reduced while chironomid densities increased. This difference may be the result of combined affects of low flows and anchor ice formation. Decreases observed in the density of EPT taxa may be the result of a scouring process that results from anchor ice formation. Many of the EPT taxa use microhabitat (surface areas) that would be susceptible to this scouring process, whereas the habitat used by most chironomids (subsurface) provides better protection. Thermograph data from December 2002, January 2003, and February 2003 identified periods of anchor ice formation immediately upstream from the FPR-TC site (Figures 11-13). The lack of water temperature data for the previous winter season (2001-2002) at FPR-TC makes it impossible to compare anchor ice formation; however, anchor ice were observed during that time period.



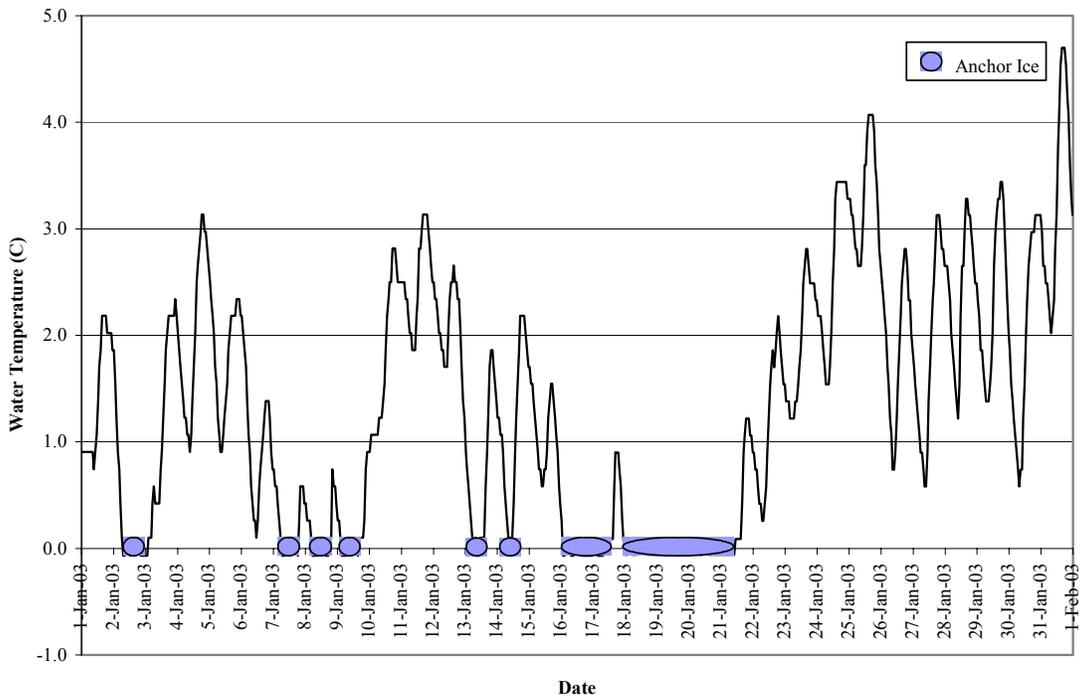
**Figure 9. Winter discharge (December-February) for the Fryingpan River below Ruedi Reservoir, Colorado.**



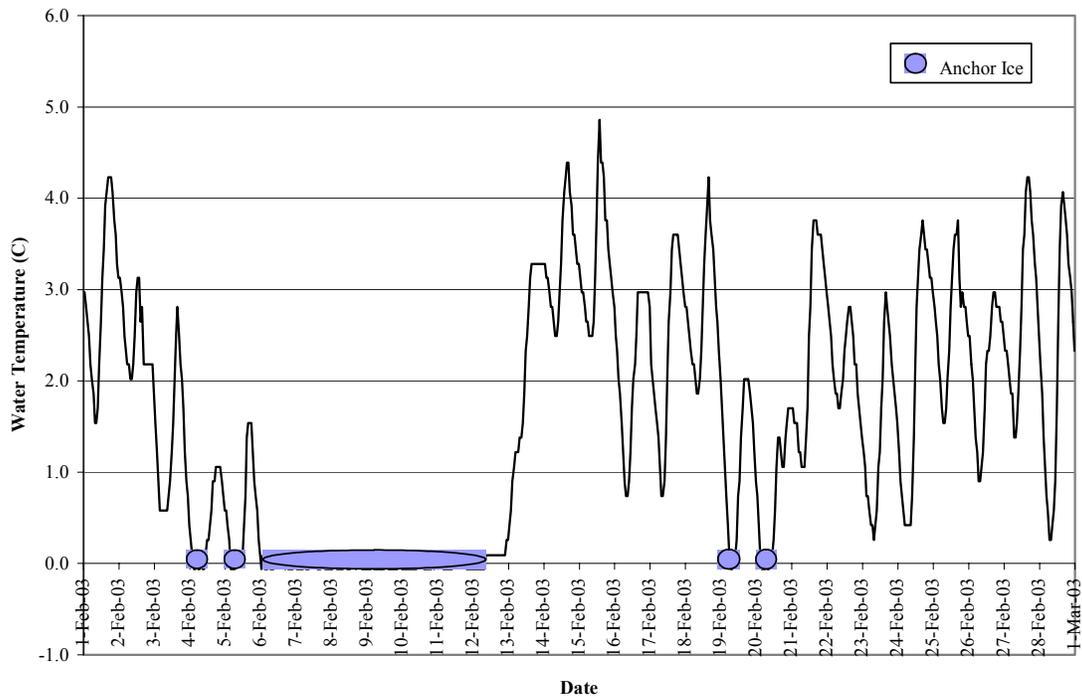
**Figure 10. Winter water temperatures for Fryingpan River below Ruedi Reservoir, Colorado.**



**Figure 11. Hourly water temperatures during December 2002 at Palm Site.**



**Figure 12. Hourly water temperatures during January 2003 at Palm Site.**



**Figure 13. Hourly water temperatures during February 2003 at Palm Site.**

The lower discharge at site FPR-TC in 2003 should have been much more conducive to the formation of anchor ice than the higher flows during the previous winter. Lower flow would be cooled more rapidly than at higher flows. However, air temperature data indicates that the 2002-2003 winter was milder than the previous winter (Table 3). If anchor ice formation is at least partially responsible for the degraded conditions observed at site FPR-TC during the spring of 2003, then conditions could have potentially been worse with colder air temperatures during the winter months.

**Table 3. Mean monthly air temperatures (°F) taken at Meredith, Colorado.**

	Mean Maximum	Mean Minimum	Mean Average
December 2001	31.1	-1.2	14.9
January 2002	29.0	-1.9	13.5
February 2002	29.1	-6.9	11.1
December 2002	35.5	6.5	21.0
January 2003	37.5	11.3	24.4
February 2003	29.2	8.1	18.7

## RECOMMENDATIONS

Questions for future studies include:

- Are benthic macroinvertebrates more influenced during the winter by changes in discharge or anchor ice formation?
- What is the specific relationship between anchor ice formation, discharge and air temperature at site FPR-TC (or other locations in the Fryingpan River)?
- In a winter with average air temperatures and cloud cover, what is the discharge that would be necessary to maintain high densities of macroinvertebrates.
- What is the time and extent of recovery that can be expected after impact to macroinvertebrate communities?

To answer the preceding questions, macroinvertebrate sampling and air and water temperature monitoring should continue in the spring and fall for at least the next year. Ideally, it would be good to compare sampling before and after winters with different base lows and different air temperature patterns. Sites for temperature monitoring and macroinvertebrate sampling should at least include FPR-RES and FPR-TC, and could be expanded if there are other areas of concern. Water quality data (collected frequently) from this reach of the Fryingpan River would also be useful.

## LITERATURE CITED

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## **APPENDIX A**

**Table 1. Macroinvertebrate data collected from the Fryingpan River at site FPR-RES on 12 October 2002.**

Fryingpan River										
FPR-RES		Sample		Mean	ni*LOGni	Count	TV		FBI	
12 Oct. 02	1	2	3							
<i>Acentrella insignificans</i>	1	9	12	7.33	6.35	1	4		0.0120	
<i>Baetis (flavistriga)</i>										
<i>Baetis (tricaudatus)</i>	279	401	200	293.33	723.76	1	4		0.4785	
<i>Drunella grandis</i>	3	1	4	2.67	1.14	1	1		0.0011	
<i>Drunella coloradensis</i>										
<i>Drunella doddsi</i>	3	7	9	6.33	5.08	1	1		0.0026	
<i>Ephemerella</i> sp.	44	36	29	36.33	56.69	1	1		0.0148	
<i>Cinygmula</i> sp.										
<i>Epeorus longimanus</i>										
<i>Rhithrogena</i> sp.										
<i>Paraleptophlebia</i> sp.	2	5	2	3.00	1.43	1	2		0.0024	
<i>Tricorythodes minutus</i>										
<i>Caenis</i> sp.										
<i>Pteronarcella badia</i>										
<i>Capnia</i> sp.										
<i>Zapada</i> sp.		1		0.33	-0.16	1	2		0.0003	
<i>Paraperla frontalis</i>										
<i>Sweltsa</i> sp.										
<i>Triznaka signata</i>										
<i>Claassenia sabulosa</i>										
<i>Hesperoperla pacifica</i>										
<i>Skwala americana</i>										
<i>Isoperla fulva</i>										
<i>Isoperla</i> sp. 2			1	0.33	-0.16	1	2		0.0003	
<i>Brachycentrus americanus</i>	1			0.33	-0.16	1	1		0.0001	
<i>Brachycentrus occidentalis</i>										
<i>Agapetus boulderensis</i>										
<i>Culoptila</i> sp.										
<i>Glossosoma</i> sp.		1		0.33	-0.16	1	0		0.0000	
<i>Arctopsyche grandis</i>										
<i>Hydropsyche cockerelli</i>										
<i>Hydropsyche occidentalis</i>			1	0.33	-0.16	1	4		0.0005	
<i>Hydropsyche</i> sp. (oslari)										
<i>Hydroptila</i> sp.	6	4	1	3.67	2.07	1	1		0.0015	
<i>Lepidostoma</i> sp.										
<i>Ceraclea</i> sp.										
<i>Oecetis</i> sp.										
<i>Dolophilodes aequalis</i>										
<i>Rhyacophila brunnea</i>	1			0.33	-0.16	1	0		0.0000	
<i>Rhyacophila coloradensis</i>		1		0.33	-0.16	1	0		0.0000	
<i>Neothremma alicia</i>										
<i>Oligophlebodes minuta</i>										
Orthocladiinae	1582	1026	512	1040.00	3137.71	1	6		2.5445	
Tanypodinae										
Tanytarsini	6	11	6	7.67	6.78	1	6		0.0188	
Chironomini		1		0.33	-0.16	1	8		0.0011	
Diamesinae		1		0.33	-0.16	1	6		0.0008	
<i>Simulium</i> sp.	50	283	58	130.33	275.66	1	6		0.3189	
<i>Protanyderus margarita</i>										
<i>Chelifera</i> sp.										
<i>Clinocera</i> sp.										
<i>Hemerodromia</i> sp.										
<i>Antocha</i> sp.	21	20	11	17.33	21.47	1	3		0.0212	
<i>Dicranota</i> sp.										
<i>Hexatoma</i> sp.										
<i>Atherix pachypus</i>										
<i>Pericoma</i> sp.		1		0.33	-0.16	1	10		0.0014	
<i>Optioservus</i> sp.										
<i>Heterlimnius corpulentus</i>	20	5	10	11.67	12.45	1	4		0.0190	
<i>Zaitzevia parvula</i>	1			0.33	-0.16	1	4		0.0005	
<i>Microcylloepus</i> sp.										
<i>Narpus concolor</i>										
<i>Hydracarina</i> sp.	8	8	2	6.00	4.67	1	8		0.0196	
<i>Gammarus</i> sp.	3	1		1.33	0.17	1	4		0.0022	
<i>Physa</i> sp.		1		0.33	-0.16	1	8		0.0011	
<i>Pisidium</i> sp.	1		1	0.67	-0.12	1	8		0.0022	
<i>Dugesia</i> sp.										
<i>Polycelis coronata</i>	336	611	655	534.00	1456.51	1	8		1.7420	
<i>Oligochaeta</i>	372	552	106	343.33	870.60	1	10		1.4000	
Nematoda	2	4	4	3.33	1.74	1	10		0.0136	
Totals	2742.0	2991.0	1624.0	2452.33	6582.25	30				6.62
Shannon Weaver Diversity					2.34					
Shannon Weaver Evenness					0.478					

**Table 2. Macroinvertebrate data collected from the Fryingpan River at site FPR-TC on 12 October 2002.**

Fryingpan River										
FPR-TC		Sample			Mean	ni*LOGni	Count	TV		FBI
12 Oct. 02	1	2	3							
<i>Acentrella insignificans</i>										
<i>Baetis (flavistriga)</i>										
<i>Baetis (tricaudatus)</i>	483	447	448	459.33	1222.80	1	4			1.2061
<i>Drunella grandis</i>	17	9	6	10.67	10.97	1	1			0.0070
<i>Drunella coloradensis</i>										
<i>Drunella doddsi</i>	1	2	3	2.00	0.60	1	1			0.0013
<i>Ephemerella</i> sp.	10	3	13	8.67	8.13	1	1			0.0057
<i>Cinygmula</i> sp.	8	5	19	10.67	10.97	1	4			0.0280
<i>Epeorus longimanus</i>										
<i>Rhithrogena</i> sp.										
<i>Paraleptophlebia</i> sp.	47	56	99	67.33	123.10	1	2			0.0884
<i>Tricorythodes minutus</i>										
<i>Caenis</i> sp.										
<i>Pteronarcella badia</i>										
<i>Capnia</i> sp.										
<i>Zapada</i> sp.										
<i>Paraperla frontalis</i>										
<i>Sweltsa</i> sp.										
<i>Triznaka signata</i>										
<i>Claassenia sabulosa</i>										
<i>Hesperoperla pacifica</i>	2	2	3	2.33	0.86	1	1			0.0015
<i>Skwala americana</i>										
<i>Isoperla fulva</i>	4	2	6	4.00	2.41	1	2			0.0053
<i>Isoperla</i> sp. 2										
<i>Brachycentrus americanus</i>	53	14	26	31.00	46.23	1	1			0.0204
<i>Brachycentrus occidentalis</i>										
<i>Agapetus boulderensis</i>		1	1	0.67	-0.12	1	0			0.0000
<i>Culoptila</i> sp.										
<i>Glossosoma</i> sp.		1		0.33	-0.16	1	0			0.0000
<i>Arctopsyche grandis</i>	12	14	9	11.67	12.45	1	4			0.0306
<i>Hydropsyche cockerelli</i>	1			0.33	-0.16	1	4			0.0009
<i>Hydropsyche occidentalis</i>										
<i>Hydropsyche</i> sp. (oslari)	3	4		2.33	0.86	1	4			0.0061
<i>Hydroptila</i> sp.	1			0.33	-0.16	1	1			0.0002
<i>Lepidostoma</i> sp.	236	149	139	174.67	391.64	1	4			0.4586
<i>Ceraclea</i> sp.										
<i>Oecetis</i> sp.										
<i>Dolophilodes aequalis</i>	1			0.33	-0.16	1	3			0.0007
<i>Rhyacophila brunnea</i>	3	2	2	2.33	0.86	1	0			0.0000
<i>Rhyacophila coloradensis</i>										
<i>Neothremma alicia</i>										
<i>Oligophlebodes minuta</i>	22	16	17	18.33	23.16	1	4			0.0481
Orthocladiinae	92	88	255	145.00	313.40	1	6			0.5711
Tanypodinae	7	3	3	4.33	2.76	1	6			0.0171
Tanytarsini	10	14	12	12.00	12.95	1	6			0.0473
Chironomini										
Diamesinae			2	0.67	-0.12	1	6			0.0026
<i>Simulium</i> sp.	2	4	6	4.00	2.41	1	6			0.0158
<i>Protanyderus margarita</i>										
<i>Chelifera</i> sp.	6	3	1	3.33	1.74	1	6			0.0131
<i>Clinocera</i> sp.										
<i>Hemerodromia</i> sp.										
<i>Antocha</i> sp.	41	14	22	25.67	36.17	1	3			0.0505
<i>Dicranota</i> sp.	1			0.33	-0.16	1	3			0.0007
<i>Hexatoma</i> sp.										
<i>Atherix pachypus</i>		1		0.33	-0.16	1	2			0.0004
<i>Pericoma</i> sp.	1	1		0.67	-0.12	1	10			0.0044
<i>Optioservus</i> sp.	1			0.33	-0.16	1	4			0.0009
<i>Heterolimnius corpulentus</i>	76	91	87	84.67	163.21	1	4			0.2223
<i>Zaitzevia parvula</i>										
<i>Microcylloepus</i> sp.			1	0.33	-0.16	1	4			0.0009
<i>Narpus concolor</i>										
<i>Hydracarina</i> sp.	16	8	20	14.67	17.11	1	8			0.0770
<i>Gammarus</i> sp.										
<i>Physa</i> sp.			2	0.67	-0.12	1	8			0.0035
<i>Pisidium</i> sp.	50	141	70	87.00	168.74	1	8			0.4569
<i>Dugesia</i> sp.										
<i>Polycelis coronata</i>	189	269	238	232.00	548.79	1	8			1.2184
<i>Oligochaeta</i>	147	61	74	94.00	185.47	1	10			0.6171
Nematoda	5	6	7	6.00	4.67	1	10			0.0394
Totals	1548.0	1431.0	1591.0	1523.33	3310.71	38				5.27
Shannon Weaver Diversity					3.35					
Shannon Weaver Evenness					0.639					

**Table 3. Macroinvertebrate data collected from the Fryingpan River at site FPR-RES on 29 April 2003.**

Fryingpan River									
FPR-RES		Sample			Mean	ni*LOGni	Count	TV	FBI
29 Apr. 03	1	2	3						
<i>Acentrella insignificans</i>									
<i>Baetis (flavistriga)</i>									
<i>Baetis (tricaudatus)</i>	177	340	255	257.33	620.30	1	4		0.4701
<i>Drunella grandis</i>	2	3		1.67	0.37	1	1		0.0008
<i>Drunella coloradensis</i>	2			0.67	-0.12	1	1		0.0003
<i>Drunella doddsi</i>		5	2	2.33	0.86	1	1		0.0011
<i>Ephemerella</i> sp.	15	45	51	37.00	58.02	1	1		0.0169
<i>Serratella tibialis</i>									
<i>Cinygmula</i> sp.	4	5	4	4.33	2.76	1	4		0.0079
<i>Epeorus longimanus</i>		3	4	2.33	0.86	1	4		0.0043
<i>Rhithrogena</i> sp.									
<i>Paraleptophlebia</i> sp.	4	7	3	4.67	3.12	1	2		0.0043
<i>Tricorythodes minutus</i>									
<i>Pteronarcella badia</i>									
<i>Prostoia besametsa</i>									
<i>Triznaka signata</i>									
<i>Sweltsa</i> sp.									
<i>Claassenia sabulosa</i>									
<i>Hesperoperla pacifica</i>									
<i>Isoperla fulva</i>									
<i>Isoperla</i> sp. 2									
<i>Skwala americana</i>									
<i>Brachycentrus americanus</i>									
<i>Brachycentrus occidentalis</i>									
<i>Micrasema bacro</i>									
<i>Culoptila</i> sp.									
<i>Glossosoma</i> sp.									
<i>Arctopsyche grandis</i>									
<i>Hydropsyche cockerelli</i>									
<i>Hydropsyche occidentalis</i>									
<i>Hydropsyche</i> sp. (oslari)									
<i>Hydroptila</i> sp.									
<i>Lepidostoma</i> sp.									
<i>Ceraclea</i> sp.									
<i>Oecetis</i> sp.									
<i>Rhyacophila brunnea</i>									
<i>Rhyacophila coloradensis</i>	1			0.33	-0.16	1	0		0.0000
<i>Neothremma alicia</i>									
<i>Oligophlebodes minuta</i>									
Orthocladiinae	1165	1223	858	1082.00	3283.03	1	6		2.9648
Tanyptodinae									
Tanytarsini	3	1	4	2.67	1.14	1	6		0.0073
Chironomini			1	0.33	-0.16	1	8		0.0012
Diamesinae	596	585	459	546.67	1496.62	1	6		1.4979
<i>Simulium</i> sp.	2	11		4.33	2.76	1	6		0.0119
<i>Protanyderus margarita</i>									
<i>Bibiocephala grandis</i>									
<i>Chelifera</i> sp.									
<i>Clinocera</i> sp.									
<i>Hemerodromia</i> sp.									
<i>Tipula</i> sp.									
<i>Antocha</i> sp.	2	1		1.00	0.00	1	3		0.0014
<i>Dicranota</i> sp.									
<i>Hexatoma</i> sp.									
<i>Atherix pachypus</i>									
<i>Pericoma</i> sp.									
<i>Optioservus</i> sp.									
<i>Heterolimnius corpulentus</i>	1	2	4	2.33	0.86	1	4		0.0043
<i>Zaitzevia parvula</i>									
<i>Narpus concolor</i>									
<i>Hydracarina</i> sp.	7	11	20	12.67	13.97	1	8		0.0463
<i>Gammarus</i> sp.									
<i>Physa</i> sp.									
Planorbidae									
<i>Pisidium</i> sp.	1			0.33	-0.16	1	8		0.0012
<i>Dugesia</i> sp.									
<i>Polycelis coronata</i>	145	139	294	192.67	440.21	1	8		0.7039
Oligochaeta	53	20	29	34.00	52.07	1	10		0.1553
Nematoda									
Totals	2180.0	2401.0	1988.0	2189.67	5976.35	20			
Shannon Weaver Diversity					2.03				
Shannon Weaver Evenness					0.470				

**Table 4. Macroinvertebrate data collected from the Fryingpan River at site FPR-TC on 29 April 2003.**

Fryingpan River										
FPR-TC		Sample			Mean	ni*LOGni	Count	TV		FBI
29 Apr. 03	1	2	3							
<i>Acentrella insignificans</i>										
<i>Baetis (flavistriga)</i>										
<i>Baetis (tricaudatus)</i>	109	263	37	136.33	291.02	1	4			0.2993
<i>Drunella grandis</i>	2	2	2	2.00	0.60	1	1			0.0011
<i>Drunella coloradensis</i>										
<i>Drunella doddsi</i>										
<i>Ephemerella</i> sp.		2	8	3.33	1.74	1	1			0.0018
<i>Serratella tibialis</i>	12	18		10.00	10.00	1	1			0.0055
<i>Cinygmula</i> sp.	18	8	14	13.33	15.00	1	4			0.0293
<i>Epeorus longimanus</i>	3	18	1	7.33	6.35	1	4			0.0161
<i>Rhithrogena</i> sp.										
<i>Paraleptophlebia</i> sp.	10	44	45	33.00	50.11	1	2			0.0362
<i>Tricorythodes minutus</i>										
<i>Pteronarcella badia</i>										
<i>Prostoia besametsa</i>										
<i>Triznaka signata</i>										
<i>Sweltsa</i> sp.										
<i>Claassenia sabulosa</i>			3	1.00	0.00	1	1			0.0005
<i>Hesperoperla pacifica</i>	4	1	3	2.67	1.14	1	1			0.0015
<i>Isoperla fulva</i>		1	1	0.67	-0.12	1	2			0.0007
<i>Isoperla</i> sp. 2										
<i>Skwala americana</i>										
<i>Brachycentrus americanus</i>	2	13	8	7.67	6.78	1	1			0.0042
<i>Brachycentrus occidentalis</i>										
<i>Micrasema bactro</i>		1		0.33	-0.16	1	1			0.0002
<i>Culoptila</i> sp.										
<i>Glossosoma</i> sp.										
<i>Arctopsyche grandis</i>	8	6	6	6.67	5.49	1	4			0.0146
<i>Hydropsyche cockerelli</i>										
<i>Hydropsyche occidentalis</i>										
<i>Hydropsyche</i> sp. (oslari)										
<i>Hydroptila</i> sp.	1			0.33	-0.16	1	1			0.0002
<i>Lepidostoma</i> sp.	23	23	44	30.00	44.31	1	4			0.0658
<i>Ceraclea</i> sp.										
<i>Oecetis</i> sp.										
<i>Rhyacophila brunnea</i>	1		2	1.00	0.00	1	0			0.0000
<i>Rhyacophila coloradensis</i>	1			0.33	-0.16	1	0			0.0000
<i>Neothremma alicia</i>										
<i>Oligophlebodes minuta</i>	4	5	5	4.67	3.12	1	4			0.0102
Orthocladiinae	901	1460	1495	1285.33	3996.12	1	6			4.2319
Tanypodinae	8	14	22	14.67	17.11	1	6			0.0483
Tanytarsini	53	103	211	122.33	255.38	1	6			0.4028
Chironomini										
Diamesinae	12	8	6	8.67	8.13	1	6			0.0285
<i>Simulium</i> sp.										
<i>Protanyderus margarita</i>										
<i>Bibiocephala grandis</i>										
<i>Chelifera</i> sp.		3	6	3.00	1.43	1	6			0.0099
<i>Clinocera</i> sp.		1	1	0.67	-0.12	1	6			0.0022
<i>Hemerodromia</i> sp.										
<i>Tipula</i> sp.										
<i>Antocha</i> sp.	6	11	21	12.67	13.97	1	3			0.0209
<i>Dicranota</i> sp.										
<i>Hexatoma</i> sp.										
<i>Atherix pachypus</i>										
<i>Pericoma</i> sp.										
<i>Optioservus</i> sp.			4	1.33	0.17	1	4			0.0029
<i>Heterolimnius corpulentus</i>	20	25	76	40.33	64.76	1	4			0.0885
<i>Zaitzevia parvula</i>										
<i>Narpus concolor</i>										
<i>Hydracarina</i> sp.	4	15	16	11.67	12.45	1	8			0.0512
<i>Gammarus</i> sp.										
<i>Physa</i> sp.										
Planorbidae										
<i>Pisidium</i> sp.	10	14	21	15.00	17.64	1	8			0.0658
<i>Dugesia</i> sp.										
<i>Polycelis coronata</i>	17	52	15	28.00	40.52	1	8			0.1229
Oligochaeta	24	21	3	16.00	19.27	1	10			0.0878
Nematoda		2	4	2.00	0.60	1	10			0.0110
Totals	1253.0	2134.0	2080.0	1822.33	4882.49	32				5.66
Shannon Weaver Diversity					1.93					
Shannon Weaver Evenness					0.386					